

Investigation of the redox-conditions in the Nam Du well field, Hanoi, Vietnam, and their implications for mobilisation of Arsenic

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Abstract

In 2005, a new well field along the Red River started pumping groundwater for production of drinking water in the Nam Du area south of Hanoi, Vietnam. Elevated concentrations of ammonium have been a well-known problem in the southern part of Hanoi area, but recently elevated concentrations of arsenic in the groundwater has also been of concern. The first study on elevated concentrations of arsenic in the Red River delta was published in 2001 (Berg et al., 2001). Peat has been discussed as a source for both ammonium and as a driver for the mobilisation of arsenic and this study aims to investigate the possible relationship between arsenic, ammonium, the peat and the redox conditions. Chemical analyses from this study resulted in total arsenic concentrations from 27 to 312 µg/l in the groundwater samples, which can be compared to the health-based guideline of 10 µg/l. Most of the arsenic in the groundwater is present as arsenite, the more toxic specie of arsenic. The analyses also showed elevated concentrations of ammonium in the groundwater ranging from 1.8 to 34.7 mg/l. The relationships between the different redox couples were studied with Visual Minteg, which showed that the redox potentials measured in the field were a little bit higher than expected from the model. This can probably be explained by contamination of oxygen during sampling and/or measurement. The lack of sulphate in combination with the present redox conditions indicated that oxidation of arsenic bearing sulphurous minerals is not likely to be the main source of arsenic. Instead, the relationship between arsenic and HCO₃, indicates that a more probable source of the arsenic would be reductive dissolution of iron oxyhydroxides.

Sammanfattning

Hösten 2005 började man pumpa grundvatten från ett nytt brunnsfält i Nam Du-området längs Röda floden strax söder om Hanoi för dricksvattenproduktion. Förhöjda halter av ammonium är sedan länge ett känt problem i grundvattnet söder om Hanoi men på senare år har även förhöjda halter av arsenik i grundvattnet gett anledning till oro. År 2001 publicerades den första studien som visade på förhöjda halter av arsenik i Röda flodendeltat (Berg et al., 2001). Torv har diskuterats som både en källa till ammoniumet och som en pådrivande faktor för mobiliseringen av arsenik. Syftet med den här studien har varit att undersöka ett möjligt samband mellan arsenik, ammonium, torven och redox-förhållandena. Kemiska analyser av grundvattnet visade på arsenikhalter mellan 27 och 312 µg/l, vilket är långt över de riktvärden som finns för dricksvatten (10 µg/l) både i Vietnam och som WHO rekommenderar. Till största delen förekommer arseniken som arsenit, vilket är den mer toxiska jonformen av arsenik. Analyser visade även på förhöjda halter av ammonium i grundvatten, med halter från 1,8 till 34,7 mg/l. Sambandet mellan de olika redoxparen studerades med hjälp av Visual Minteg, som visade att den i fält uppmätta redoxpotentialen var något högre än vad som kunde förväntas utifrån modellen. Detta kan troligtvis förklaras av syrekontaminering av proverna under provtagning och/eller mätning. Avsaknaden av sulfat i kombination med rådande redoxförhållanden visar att oxidation av svavelinnehållande mineral inte är en trolig källa för arseniken. Istället indikerar sambandet mellan As och HCO₃ att reduktion av järnhydroxider är en mer trolig källa till arseniken.

Tóm tắt

Năm 2005, một bãi giêng mới nằm dọc sông Hồng - thuộc địa bàn khu vực Nam Dự, phía Nam Hà Nội, Việt Nam - bắt đầu đi vào hoat động phục vụ cung cấp nước cho ăn uống, sinh hoạt. Mức độ nhiễm bẩn Amonii ở khu vực phía Nam Hà Nội đã trở nên báo động. Bên canh đó, nhiêm bân Asenic trong nước dưới đất đang là vân đề cân phải quan tâm. Công trình nghiên cứu đầu tiên về mức độ nhiễm bẩn Asenic ở đồng bằng sông Hồng đã được công bố năm 2001 (Berg & nnk., 2001). Than bùn chứa trong các lớp trầm tích đã được xem xét như là một nguồn gây ô nhiễm Amoni và nguyên nhân tạo ra khả năng di chuyển Asenic vào trong nước dưới đất và nghiên cứu này nhằm mục đích điều tra khả năng giữa các nguồn nhiễm bẩn Asenic, Amonii và điều kiên ô xy hóa - khử. Phân tích thành phần hóa học các mẫu nước dưới đất cho thấy hàm lượng Asenic tổng biến đối từ 27 đến 312 µg/l cao hơn tiêu chuẩn nước ăn uống - sinh hoat là 10µg/l. Phần lớn dang tồn tại của Asenic trong nước dưới đất ở đây là Asenicite, một dạng có khả năng độc tố cao của Asenic. Kết quả phân tích cũng cho thấy mức độ nhiễm bẩn của Amonii trong nước dưới đất biến đổi từ 1.8 đến 34.7 mg/l. Mối quan hệ giữa các cặp ô xy hóa - khử khác nhau được nghiên cứu trên phần mềm Visual Minteq, mà ở đó cho thầy thế ô xy hóa - khử đo đạc hiện trường cao hơn một chút so với mô hình. Điều này có thể giải thích rằng có thể ô xy đã đi vào trong nước trong quá trình lấy mẫu và đo đạc ngay tại hiện trường. Hàm lượng Sulphat nhỏ trong mối liên hệ với điều kiện ô xy hóa - khử chỉ ra rằng quá trình ô xy hóa Asenic do khoáng chất Sulphua giường như không thể là nguồn hình thành chính của Asenic. Thay vào đó, mối quan hệ giữa Asenic và HCO₃ chỉ ra rằng nguồn hình thành Asenicic chủ yếu có thể là quá trình khử hydroxit sắt.

Preface by the supervisors

High concentrations of arsenic in groundwater have been reported from different regions in the world. Well-known high-As groundwater areas have been found in Argentina, Chile, Mexico, China and Hungary, and more recently in the densely populated river deltas in S and SE Asia which contain large groundwater aquifers: West Bengal (India), Bangladesh and Vietnam. The most common cause of the high-As groundwater in these regions is due to natural occurrence of arsenic in the sediments and the right conditions for arsenic mobilisation.

The first publication on arsenic contamination of the aquifers in the densely populated Red River delta was published in 2001. Consequently, in recent years, a major concern regarding the arsenic concentration in the groundwater in the Red River delta has arisen. The process for mobilisation of arsenic in the Red River delta aquifers is believed to be due to the natural conditions. However, in order to manage the groundwater quality in the future, a deeper understanding of the prevailing conditions and the on-going processes is needed. Furthermore, the Hanoi aquifers in the Red River delta have high and increasing ammonium concentrations, of which the source, so far, have been undetermined.

The work presented in this study has been an important part of a current research project at the Swedish Geotechnical Institute (SGI): *Sources and distribution of ammonium and arsenic in the Red River delta aquifers* (AmonAs). AmonAs is financed by Sida/SAREC during the period 2005 – 2007 and is a co-operation between SGI, the Royal Institute of Technology (KTH) and Hanoi University of Mining and Geology (HUMG) in Vietnam.

Johanna Moreskog has made a study in order to investigate a possible relationship between the arsenic, ammonium, the peat and the redox-conditions in the groundwater in Nam Du. The chemical composition of the groundwater was analysed and used for geochemical modelling in order to investigate the redox conditions. By using Visual Minteq, both ion speciation and solubility calculations were performed. An extensive field campaign was carried out in Vietnam during February – March 2006, together with Peter Harms-Ringdahl. The results of this study support the current hypothesis that the mobilisation of arsenic is caused by reduction of iron hydroxides.

Johanna and Peter planned and carried out the field campaign in a highly independent manner and cooperated effectively with their Vietnamese co-students. Johanna has worked impressively independent with the analyses and the writing of this thesis. She has ambitiously widened the scope of her work and her knowledge during the progress of this study.

It has been our pleasure to have the opportunity to work together with her.

We wish you all the best in the future!

Göteborg 2007-08-16

Jenny Norrman, SGIHåkan Rosqvist, NSR ABGunnar Jacks, KTHPhD, supervisorPhD, supervisorProf Emeritus, supervisor

Abbreviations

Equipment

AAS – Atomic Absorptions Spectrophotometry HG-AAS – Hydride Generation Atomic Absorptions Spectrophotometry IC – Ion Chromatograph ICP-MS – Inductivley Coupled Plasma – Mass Spectrometer ICP-OES – Inductively Coupled Plasma – Optical Emission Spectrometer MS – Mass Spectrometer VM – Visual Minteq, Geochemical modelling software

Universities, laboratories, institutes etc

CETASD – Centre for Environmental Technology and Sustainable Development
EAWAG –Swiss Federal Institute of Aquatic Science and Technology
HUMG – Hanoi University of Mining and Geology
HUS – Hanoi University of Science
INST – Institute of Nuclear Science and Technology
SGI – Swedish Geotechnical Institute
SLU – Swedish Agricultural University
SU – Stockholm University
UMU – Umeå University
WHO – World Health Organisation
ÅAU – Åbo Akademi University

Codes for sampling sites

1,2,3 - Transect number
RGw - Shallow groundwater close to the river
SRw - Surface water from the river
D - Surface water from ponds
P1, P3, P86, ND12 - Monitoring wells
PX, PY - Private, small scale production wells
ND 3, ND 9 - Big scale production wells

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It was also of great value for the understanding and evaluation of laboratory anlyses to be able to visit the labs in Vietnam, INST and CETASD, many thanks to the helpful staff at these laboratories. Michael Berg at EAWAG, has also been of great help by kindly answering all questions and analysing our water samples.

This Bachelor's thesis is examined at the Department of Soil Sciences at the Swedish University of Agricultural Science (SLU) and the employees, especially at the Division of Soil Chemistry and Pedology, are gratefully acknowledged for answering questions at all times.

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1. Introduction

1.1. Background

High concentrations of arsenic has earlier been found in aquifers in Bangladesh, Argentina, China, Mexico etc, (WHO, 2001; Smedley and Kinniburgh, 2002; Bhattacharya et al., 2006) and in 2001, Berg et al. published the first report on the high concentrations of As in the Red River delta in northern Vietnam (see Figure 1-1). The average concentration was 159 μ g/l and peak values of 3050 μ g/l were observed, these values are well above the international limits of arsenic in drinking water set to 10 μ g/l by WHO (Berg et al., 2001). In reducing environments, it is typically groundwater in late Quaternary sediments, mainly of Holocene origin, of deltaic and alluvial origin that might contain elevated concentrations of arsenic, which is the situation in both Bangladesh and Vietnam (Berg et al., 2007). It is also suggested by Saunders et al. (2004), that similar aquifers in developed countries also may contain elevated concentrations of arsenic. The reason for this not being noted yet would be the low usage of this type of groundwater due to the high iron content. Even small changes in the environment can mobilise the arsenic, and due to the low guideline, have severe consequences (WHO, 2001).



Figure 1-1. Map of arsenic, ammonium and peat in the area around Hanoi in the Red River delta, Vietnam.

The consequence of long-term exposure (10-15 years minimum), of arsenic can be e.g. bladder and skin cancer, black foot disease (WHO, 2001). In a field campaign in 2001, Agusa et al. found levels of arsenic in hair from people in two districts south of Hanoi (Thanh Tri and Gia Lam) that exceeds the levels where symptoms appear (Agusa et al., 2006). Many of the symptoms of chronic arsenic poisoning takes long time to develop, which is why present status of arsenic poisoning just might be the tip of an ice berg (van Geen et al., 2004).

Southeast of the city (see Figure 1-1), where some of the highest concentrations were measured, a new well field (Nam Du well field) were built and started producing drinking water in 2005. In the area, also elevated levels of ammonium are previously noted. (Nhan, 2006; Berg et al., 2001)

In Bangladesh elevated levels of arsenic is a enormous problem and the conditions are similar to those in Vietnam. Peat is present in the sediments in the Ganges delta in Bangladesh, which is believed to initiate the reductive processes that are causing the mobilisation of arsenic in Bangladesh. Also in the Red River delta in Vietnam, peat layers are present (see Figure 1-1) and, for the same reasons, they are believed to contribute to the mobilisation of arsenic. (Berg et al., 2001; McArthur et al., 2004)

Elevated levels of ammonium are normally not found in surface water, since it will oxidise to nitrate. However, elevated levels exceeding the national guidelines are noted south of Hanoi, where the guideline is set to 1.5 mg/l (Nhan, 2006). Andersson and Norrman (1998) suggested three sources to the elevated concentrations of ammonium in the aquifers of Hanoi. These are infiltration from Red River, degradations of peat and surface activities, e.g. fertilizers, sewage water etc. By making a nitrogen budget of the area, the first source could be excluded, since the nitrogen concentrations in the Red River not could cause such levels of nitrogen in the groundwater.

In Bangladesh, which in many ways has similar conditions as the Red River delta, the most widely accepted theory of arsenic mobilisation is the reduction of iron oxyhydroxides. When organic matter is microbially oxidised, the arsenic bearing iron oxyhydroxide is reduced and dissolved, and consequently the arsenic is mobilised (Smedley and Kinniburgh, 2002; Anawar et al., 2003). Therefore to investigate the redox conditions is one approach to understand the mobilisation of arsenic also in the Red River delta.

1.2. *Objectives and scope*

The redox condition is an important factor governing the mobilisation of both arsenic and ammonium. Therefore, the objective of this study is to investigate the present redox processes in the Nam Du well field.

This was done by

- measuring redox and different species during the field campaign,
- to investigate the relationship between redox and concentrations of arsenic and ammonium
- to model the redox conditions and to study the redox sequence and mineral solubility in Visual Minteq, a geochemical modelling software.

1.3. Outline of the project

The project started with literature studies of the field site and the surrounding areas, followed by a field campaign in Hanoi, Vietnam. The field campaign was carried out by Johanna Moreskog and Peter Harms-Ringdahl, during the period February – March 2006, together with Vietnamese students and supervisors. The focus of the field campaign was on the spatial distribution of arsenic and ammonium in the Nam Du area. A number of sampling points were identified for both surface- and groundwater, to form three transects perpendicular to the river. At one location on the middle transect, a drilling was preformed to collect sediment samples.

The samples from groundwater, surface water and the sediments were sent to three different laboratories for analyses. Based on the results of the analyse, the redox conditions were modelled in Visual Minteq, a geochemical model software.

The thesis is a part of the AmonAs project, a Swedish-Vietnamese cooperation funded by Department for Research Co-operation (SAREC) at Swedish International Development Cooperation Agency (SIDA). The aim is to understand the spatial and temporal distribution of ammonium and arsenic in the Nam Du well field in Vietnam. The Swedish part of the project involves the Swedish Geotechnical Institute (SGI) and the Royal Institute of Technology (KTH). The Vietnamese part of the project consists of the Department of Hydrogeology at Hanoi University of Mining and Geology (HUMG). The AmonAs project is divided into two separate field campaigns, of which this has been the first.

The results of the field campaign are partly presented in this Bachelor's thesis, but also in the Master's thesis by Peter Harms-Ringdahl (Harms-Ringdahl, 2007). Since the field campaign was performed by both students, some parts of the reports are identical. Chapter 2 (Study area, Nam Du well field) is mainly written by Johanna Moreskog and Chapter 4 (Methods and material) is mainly written by Peter Harms-Ringdahl.

Finally, the results and conclusions were compiled into this Bachelor's thesis, examined at The instution of soil chemistry and pedology at the Swedish Agricultural University in Uppsala, Sweden.

2. Study area: Nam Du well field

2.1. Geography and climate

<u>Hanoi</u>

Hanoi, the capital of the Socialist republic of Vietnam, is situated in the centre of the 11 000 km² Red River delta on the Bac Bo plain in northern Vietnam, an area with approximately 11 million inhabitants (Trafford et al., 1996), (see Figure 2-1). In 2004,



Figure 2-1. Map of Vietnam (CIA, 2007)

Hanoi was inhabited by 3.1 million people and with a growth rate of 1.5% annually, the city is expected to have around 5 million inhabitants in the year 2020 (van Horen, 2005).

In Hanoi, the treatment of sewage water is poor, liquid waste in the town is disposed through open drains which run along the edge of the streets to larger drainage canals or branches of the Red River. Every evening truck collects the solid waste, which is swept into the streets, and transports it to waste sites in the area. There are no strict border between the town and farmland and the percentage of farmland increases in the suburbs. Animal waste, urea and waste water from the town are used as fertilisers on the fields (Trafford et al., 1996).

Extraction of groundwater began about 100 years ago during the French govern (Dang et al., 1996) and since the 1960s, the extraction has increased rapidly (Trafford et al., 1996). Today about 500 000 m³/day is officially extracted (Duong et al., 2003), mainly from the Hanoi aquifer system at ten major well fields (Berg et al., 2007). The water passes the eight major water treatment plants prior to distribution (Duong et al., 2003). In addition, there is an unknown number of small private tube wells extracting

water, mainly from the upper aquifer and the total estimated extraction reaches 700 000 m^3 /day (Nhan, 2006). In 2010, the extraction is expected to reach 1,000,000 m^3 /day (Dang et al., 1996).

<u>Climate</u>

The Hanoi area has a hot and rainy season with a mean temperature of 29°C in May – October. Of the average annual precipitation of 1700 mm, most of the precipitation (80-90%) comes in July - September. The cool and dry season is November – March with a mean temperature of 16°C. In the end of the cool and dry season, drizzles and monsoons are frequent. Since the recharge is restricted to the rainy season and the annual evaporation is 1025 mm, the evaporation exceeds the precipitation during the dry season and causes a drawdown of the groundwater table (Trafford et al., 1996; van Horen, 2005).

The Red River and its delta

The area is a flat delta landscape with only small altitude differences (1.9 - 10.6 m) and the Red River floats through the landscape. The river is 1150 km long and originates in the eastern Tibetan plateau in the Yunnan province, China (Tanabe et al., 2003). It continuous in a NW-SE direction through the mountainous area in the north-west of Vietnam down to the Gulf of Tonkin in South China sea. The Red River received its name due to the iron-rich silt, which gives the river a red colour. The catchments area of the Red River is 170 000 km² and over the year the discharge varies between 700 – 23000 m³/s, with an average discharge of 3740 m³/s (Tanabe et al., 2003; Berg et al., 2007).

Nam Du well field

The Nam Du well field is located 10 km south east of Hanoi city along the Red River $(20^{\circ}58'56'' \text{ N}, 105^{\circ}55'02'' \text{ E to } 20^{\circ}57'55'' \text{ N}, 105^{\circ}53'43'' \text{ E})$ in the Thanh Tri district (Figure 2-2). The area around the well field is relatively flat, with an elevation around 6.5 – 11 m above sea level.



Figure 2-2. Map over the Hanoi area. The Nam Du well field is situated south east of the town centre.

There is a dyke with a road running along the river where the nine production wells, with a total capacity of 30 000 m³/day, are located. Water from the well field passes a treatment plant before distributed to the citizens of Hanoi. The production wells distance from the river depends on the seasons but for the time of sampling (dry period) they were located approximately 200 m from the river, with 250 m between each well. A second well field was under construction during the time of the sampling campaign, Nam Du 2, and in the autumn 2006 some of the wells were started to test the water production. Nam Du 2 is a continuation of the Nam Du 1, south along the river, with nine more wells and the same total production capacity as ND 1, 30,000 m³ per day. ND 2 is located in the same direction as Nam Du phase 1, continuing south along the Red River. A third phase is being planned south of the second phase. The reason for locating the wells along the red river is to use the

river for increased recharge and at the same time minimise ammonium and arsenic contamination. About 50% of the recharge is coming from the Red River (Nhan, 2006).

The area of the well field is separated from the city by farmland and there are several small ponds in the area. The area is dominated by agricultural activities (Figure 2-3). Examples of crops are rice (paddy), bananas and vegetables. During the field measurements, it was noticed that many farmers were spraying pesticides on their crops only a few meters away from the production wells. The main industries in the area are brick and sand production, and along the second phase (see below) of the well field there is a large harbour. Brick factories were situated at the shore of ponds two and three and all ponds were surrounded by farmland (Figure 2-4).



Figure 2-3. Photo with production well ND9 to the left, surrounded by farmland.



Figure 2-4. Photo of pond 2, with a small brick factory to the left.

2.2. Geology

The geological formations in the Nam Du area are mainly of Quaternary and Pliocene origin. The sedimentological history is complex because of transgression and regression of the sea level. This has created formations with different thickness in the area. In general, most of the formations are thinnest in the NW and thickest in SE of the Red River delta. The formations might also be discontinuously distributed, and be absent at some parts (Dang et al., 1996; Trafford et al., 1996; DTU, 2005). For an overview of the geological time scale, see Figure 2-5. For an overview of the sediments profile with formations and aquifers, see Figure 2-6. If not stated other the following sections are based on a report from Dang et al. (1996).

Era	Period		Epoch	Start, million years ago
	Quartonomy		Holocene	0.11
	Quartenary		Pleistocene	1.8
	Tertiary	Naagana	Pliocene	5.3
Cenozoic		Neogene	Miocene	23
		Paleogene	Oligocene	34
			Eocene	56
			Palaeocene	65

Figure 2-5. Overview of the geological time scale under the cenozoic era, constructed after information from Tarbuck and Lutgens (2002).



Figure 2-6. Cross section of the profile with formation names, sea level change, aquifers, materials and geological epochs (Mathers and Zalasiewicz, 1999; Nghi, 2004; Dang et al., 1996).

The Holocene formations

Thai Binh (QIV3 tb)

The youngest sediments are the Thai Binh formations, of late Holocene origin, which mainly consists of alluvial deposits from the Red River and was formed from 3000 years ago until present time. The upper Holocene formation can be divided into two subgroups: upper (QIV3 tb2) and lower (QIV3 tb1) Thai Binh formation.

The upper Thai Binh formation contains mainly cobbles, pebbles, sand, clay, and silty clay with brown and grey-yellow colours. While the average thickness of the upper formation of 2.5 - 7 m, the thickness of the lower Thai Binh formation can reach 27 m. This formation consists mainly of grey medium grained sand mixed with silty sand, silty clay and layers with brown-grey humic material derived from plants. Thick layers of peat have been found in this formation (Trafford et al., 1996; Berg et al., 2001; UNICEF, 2001).

Hai Hung (QIV1-2)

The Hai Hung formations, the lower Holocene subdivisions, are formed from lacustrine, palustrine and marine deposits consisting of silty sand, silty clay and gravel mixed with peat and plant remains. These formations, with an average thickness of 1 - 10 m, are found at a depth of 23.5 - 37 m and consist mainly of sand.

The Pleistocene formations

Vin Phuc (aQIII2 vp)

The Vin Phuc formation in the Nam Du area has a mean thickness around 12 m, increasing to south east.

The upper Pleistocene formation, Vin Phuc, consists of an upper clay layer mixed with sand lenses of brown and red colours and scattered Peat formations (UNICEF, 2001). In this layer, spores and pollens grains of ferns etc. were found. The fractions in the lower formation are gravel, pebble, sand, silt, and clay of yellow –yellow-grey colour. The grains are medium to well sorted and rounded. At some locations in this layer, peat and fresh water algae can be found.

Hanoi formation (aQII-III1 hn)

The Hanoi formation is from the middle Pleistocene and consists of gravel mixed with sand of alluvial origin. The higher up in the formation, the finer is the fractions and in the top, local clay lenses can occasionally be found. The average thickness in the Nam Du area is 18 m and scattered peat is occasionally found in the top.

The uppermost part of the Hanoi formation consists of clayey, sandy silt, cobble, mixed with clay of yellow grey, yellow brown colour that originates from flood plains. Occasionally are lenses of black grey silty clay, mixed with humic material. The average thickness is 4 m. In the upper part there are traces from fresh water pollens.

In the middle of the Hanoi formations there are small pebbles, coarse sand and silt with a grey yellow colour, with an alluvial origin. The sediments are mainly composed by quartz and silica feldspar. In this part, the thickness varies between 1.5 - 7.3 m.

The bottom formation consists mainly of coarser fractions such as cobbles, pebbles and gravels, mixed with sand and silt of alluvial origin. The cobbles consist of quartz, silica and extrusive rocks. This is also the thickest part of the Hanoi formations, between 10 - 19 m.

Le Chi (aQI lc)

Below the Hanoi formation is the Le Chi formation of early Pleistocene origin. The thickness varies between 0.5 - 2.5 m, and is present at depths of 60 - 63 m in the Nam Du well field.

The upper part consists mainly of sand, silt and clay mixed with organic matter. In contrast to the middle part of the formations, the grains in the upper part are of alluvial origin and is neither well sorted nor well rounded.

In the middle part the soil fractions are mainly fine sand, silty sand of grey yellow colour, also here with an alluvial, or near alluvial, origin. The lowest part consists of a mixture of well rounded cobbles, pebbles and sand, silt and clay deposited by the mountain rivers (Dang et al., 1996; DTU, 2005).

The Pliocene formations

Vinh Bao (N2 vb)

Below the quaternary system, is the Neogene formation from the Pliocene epoch. This formation starts at a depth of 64 - 71 m and continues down approximately 100 m, but no borehole has penetrated the formation completely. This layer consists of weak conglomerates of sandstone, clay stone and well-rounded pebbles. When prospecting for the well field, fossils of organisms that lived in shallow marine environments, close to the shore were found in the Pliocene.

2.3. Hydrogeology

The hydrogeological features of the area such as permeability, water bearing capacity and hydrodynamic characteristics have been documented (Dang et al., 1996). Six major geohydrological units could be distinguished see Figure 2-6. Similarly to the geological formations, also the extent of the geohydrological formations generally increases from NW to SE (Trafford et al., 1996). Unless stated otherwise, this chapter is based on the work of Dang et al., in 1996.

- A. Aquifer in Holocene sediments, (TCN qh)
- B. Aquiclude in Pleistocene-Holocene sediments (LCN 1)
- C. Aquifer in upper Pleistocene sediments (TCN qp 2)
- D. Aquiclude in middle-upper Pleistocene sediments (LCN 2)
- E. Aquifer in lower-upper Pleistocene sediments (TCN 1 qp 1)
- F. Water bearing formations in Neogene sediments (PHCN N)
- A. <u>Aquifer in Holocene sediments (TCN qh)</u>.

In the Nam Du well field, the thickness of this aquifer varies between 4 - 34 m, while average thickness is 21 m. The groundwater table is normally found at a depth of 1 - 6 m, which varies with the landscape and throughout the year. This is a confined or locally confined aquifer and is overlain by a discontinuously distributed layer (QIV3 tb2). Pumping tests from wells in the area shows that the specific capacity (Q) is 1.06 - 20.87 l/sm and the transmissivity¹ ranges from 2.31 * $10^{-4} - 9.14 * 10^{-3}$ m²/s. Abstraction from qh is mainly from small private wells (DTU, 2005). In the Nam Du well field this aquifer has a close hydraulic connection to the Pleistocene aquifers (qp1 and qp2), and at some points there is a vertical leakage from the Holocene aquifer and downwards.

B. Aquiclude in Pleistocene-Holocene sediments (LCN 1)

The LCN 1 aquiclude is of early Holocene and late Pleistocene. The thickness varies between 1 - 16 m and the average hydraulic conductivity coefficient K, is 6.13×10^{-7} m/s in LCN 1.

C. <u>Aquifer in upper Pleistocene sediments (TCN qp 2)</u>

Aquifer qp 2 is present in the coarser fractions of the lower Vinh Phuc formation. In Nam Du, this aquifer is encountered at depths around 16 - 42 m and has an average thickness of 11 m. Q, the specific capacity, varies between 0.18 - 5.35 l/sm, and the transmissivity¹ coefficient varies from $1.85 * 10^{-4} - 5.10 * 10^{-2}$ m²/s (K $1.68 * 10^{-5} - 4.64 * 10^{-3}$ m/s), but has an average of $2.08 * 10^{-3}$ m²/s. In the Nam Du area, this aquifer is connected to the Red River. The static water level varies between 0.7 - 5.9 m

D. Aquiclude in middle-upper Pleistocene sediments (LCN 2)

The LCN 2, from the upper part of the Hanoi formation, is not distributed continuously over the Nam Du area, this creates hydraulic window between qp 1 and qp 2. The average hydraulic coefficient K is 9.26×10^{-8} m/s, much lower compared to 6.13×10^{-7} m/s in LCN 1. However, due to the hydraulic windows in the aquiclude, and there is a hydraulic contact between the Holocene and the upper Pleistocene aquifers.

E. <u>Aquifer in lower-upper Pleistocene sediments (TCN 1 qp 1)</u>

The Hanoi aquifer (qp 1) is present in materials in the lower Hanoi and the Le Chi formations, and is found at depths of 35 - 71 m. The average thickness is 21 m, but ranges from 14 - 46 m. Pumping tests from wells in the area shows a specific capacity (Q) of Q = 6.97 - 23.7 l/sm. The transmissivity¹ ranges from between $9.83 \times 10^{-3} - 3.07 \times 10^{-2}$ m²/s (K is $4.68 \times 10^{-4} - 1.46 \times 10^{-3}$ m/s). The abstraction from this aquifer has increased since the 1950's and is mainly for production wells (Trafford et al., 1996). In the Nam Du well field, qp 1 is closely connected to both qh and qp2, as well as the Red River. The static water level varies between -12 and -24 m.

F. <u>Water bearing formations in Neogene sediments (PHCN N)</u>

Directly under qp 1, at depths of 64 - 81 m, are the Neogene formations with water bearing capacity. The specific capacity, Q, is lower than in the upper aquifers, varies from 0.005 - 3.46 l/sm and the transmissivity1 is $6.37 * 10^{-4} - 9.71 * 10^{-3}$ m²/s (Dang et al., 1996). The

¹. In the reference the term transmissibility is used, but since it is stated as m^2/d and describes water flow, the author assumes it is transmissivity

static water level is 0.3 to 12.3 m. At some locations it contains brackish water (Trafford et al., 1996).

Upper and lower aquifer system

Some authors (e.g. Trafford, 1996 and DTU, 2005) describe the hydrogeological compositions of the area in terms of the upper aquifer system (UAS) and the lower aquifer system (LAS). The UAS includes the Holocene aquifer qh and the upper Pleistocene aquifer qp 1, which are separated discontinuously by the aquiclude LCN 1. The thickness of UAS varies from 10 - 45 m. Groundwater from the UAS is primary important as a source for recharge of the LAS, rather than a direct source for drinking water (Trafford et al., 1996) The LAS is the Hanoi aquifer, qp 2. LAS, like UAS increases from northwest to southeast and the thickness varies from 16 - 100 m (UNICEF, 2001).

2.4. Geological formations and mineral occurrence

This section is based entirely on information from UN, 1990. In the north Bac Bo plain, rock types such as conglomerate, sandstone, shale, chert siliceous limestone, shale with bauxite are abundant. In the north-east of the delta there are sedimentary deposits and residuals of kaolin, feldspar, bentonite, diatomite, pozzolanic clay and coal. North of the delta, barite, tin, zinc-lead and cupper of post magmatic origin are present and in the northern most districts there are large aluminium (bauxite) deposits and some zinc-lead deposits.

In the area where the Red River flows into Vietnam, phosphates are abundant, and copper, molybdenum, uranium and dolomite are present. On the way to Hanoi, the Red River passes through areas rich in iron and kaolin. West of Hanoi, there are large occurrences of post magmatic talc, asbestos and serpentine and in the western Bac Bo plain, rhyolite, orthopyre, tuff and basalt dominates.

Other minerals that are present in the northern Vietnam are Mn, Ti, Cr, Ni, Sb, Hg, W, Be, Zr, Au, Ag, V, Mo and U amongst the metallic minerals. The non-metallic minerals also present are pyrite, barite, fluorite, graphite, crystal quartz, mica, corundum, sillimanite, silica sand, quartzite gravel sand and acid, intermediate igneous rocks. Only at one spot in northern Vietnam, about 75 km north of Hanoi, a small occurrence of arsenic is present.

3. Theory

3.1. Equilibrium and solubility

A chemical equilibrium is described by the general reaction equation based on the law of mass action (Equation 3-1). The symbols A-D represents elements in solid phase, aqueous solution or in gaseous phase (e.g. $Fe^{2+}_{(aq)}$, $FeOOH_{(s)}$), and a-d represents the mole fraction. The equation is valid under ideal conditions and the constituents are expressed in concentrations. The equilibrium constant, K, is defined at a specified temperature. Thus, with knowledge of the minerals present, the general reactions and their equilibrium constants, it is possible to estimate the concentrations of ions in the surrounding solution with solubility calculations, under ideal conditions. Solubility calculations however, are expressed in activities, marked out with {X} (see Equation 3-2) even though quantities of elements normally are given as concentrations expressed with [X]. The activity is related to the concentrations by an activity coefficient (γ). (Appelo and Postma, 2005; Gustafsson et al., 2006)

aA + bB
$$\leftrightarrows$$
 cC + dD (Equation 3-1)

$$K = \frac{\{C\}^c \{D\}^d}{\{A\}^a \{B\}^b}$$
(Equation 3-2)

In reality however, there are several other factors affecting the solubility, e.g. free metal ions are forming complexes causing the concentrations of free ions to change. The fact that almost no system is in equilibrium in nature also distances this expression from reality. If the concentrations are based on measured values of the free ion concentration in water samples, K is expressed as IAP (Ion Activity Product). The ratio between the two is defining the Saturation Index (Ω or SI, see Equation 3-3).

$$\frac{IAP}{K} = SI(\Omega)$$
 (Equation 3-3)

If $\Omega > 0$, the mineral is over saturated and present in solid phase, and if $\Omega < 0$, the mineral is under saturated and is dissolved in the solution. To perform equilibrium calculations for several compounds can, however, be complicated and time demanding. For this purpose models have been developed to perform large calculations for e.g. groundwater. In this study, VisualMinteq will be used (see chapter 6)

3.2. Redox processes

A redox process is the transfer of electrons between atoms. When a substance is *reduced* it receives an electron and when it is *oxidised* it has lost an electron. While the substance giving away an electron is the *reductant*, the atom receiving the electron is the *oxidant*. However, this kind of electron exchange does not occur at any significant rate, unless it is catalysed by microbial activity. (Andersson and Norrman, 1998; Gustafsson et al., 2006) Redox reactions are often expressed in half reactions, e.g. Equation 3-4.

 $Fe_{(s)} \leftrightarrows Fe^{2+} + e^{-}$ (Equation 3-4)

This expression indicates that the electron is free in the aqueous solution, which in fact, is not a realistic assumption. In order to balance a redox reaction, a reduction and an oxidation are added to a complete redox reaction. Even though the assumption that the electron is free is unrealistic, it is a useful description of the redox conditions in the solution. In the same way as one can use the negative logarithm of the proton (pH) to describe the acidity, the negative logarithm of the electron activity (pe) can be used to describe the oxidizing ability. Unlike pH, pe can take negative values. A low pe indicates on high electron activity, and a high pe describes low electron activity. (Sposito, 1998). In order to measure the redox potential (Eh), a measuring electrode (normally made from platinum) and a reference electrode (an electrode that gives a well defined potential, stable over a longer timer e.g. Ag/AgCl₂) is needed. A voltmeter is connected between the two electrodes, which measures the electron transfer between the electrodes. (Back, 2001) The redox potential and the electron activity are related to each other according to the expression below (Appelo and Postma, 2005).

Eh (V) =
$$\frac{RT \ln 10}{F} pe = 0.059$$
 pe (Equation 3-5)

F = Farradays constant (96.42 C/mol) $R = 8.3145 J mol^{-1} K^{-1}$ T = Temperature (K)Eh = Redox potential (V)pe = electron activity

In natural environments, pe varies from -6 up to +13, under near neutral pH. For classification of oxidation states, see Table 3-1. Redox processes in groundwater normally occurs when an oxidant is added to the system, typically oxygen or nitrate but the process can also be initiated by the addition of a reductant, like dissolved organic matter (DOC). Some elements are affected more than others when redox conditions change. The six most important elements under natural conditions are C, N, O, S, Mn and Fe, and in contaminated environments, As, Se, Cr, Hg and Pb also have to be taken into account (Sposito, 1998).

Table 3-1 The definitions of oxidation states at pH 7 in terms of pe according to Sposito (1998).

Oxidation state	Range of pe at pH 7
Oxic	pe > +7
Suboxic	+2 > pe < +7
Anoxic	pe < +2

If the sediments is a closed system (e.g. a flooded sediments and abundant source of carbon) there is a well defined sequence for reduction of the, naturally occurring, principal elements (see Figure 3-1). Oxygen is the most reactive element, and already around pe +11.0 there are enough electrons to reduce O_2 to H_2O . Below pe +5.0, O_2 is no longer stable, since the oxygen that still exists will be consumed by micro-organisms. When oxygen is not longer present, micro-organisms has to find another electron acceptor than oxygen. By reducing other molecules, such as nitrate and iron oxides, it is possible to yield energy when oxygen is absent. Below pe +8.0, NO_3^- is being reduced, and below pe 5.0 and

7.0 the solid phases of manganese and iron are reduced. Further, when pe is sinking below +2, sulphate is reduced. Since different micro-organisms are reducing different elements,



Figure 3-1. The reduction sequence of the principal elements occurring naturally including arsenic.

this sequence is not only a reduction sequence, but also an ecological sequence for the activity of micro-organisms (Sposito, 1998). Since microorganisms are more efficient degrading one isotope, the micro-organism will fraction the isotope composition during its metabolism. This is important when studying the redox sequence, since the heavy ³⁴S isotope will be enriched, and ³²S will be depleted, in the residual sulphate during sulphate reduction (Dalsgaard and Bak, 1994;Clark and Fritz, 1997). Thus the isotopic composition of sulphate gives an indication of the prevailing redox conditions.

In a Pourbaix diagram, the activity of the aqueous electron (pe) and proton (pH) are shown, and under which conditions these are available. This type of diagram can also indicate the relative dominance of different redox species, see Figure 3-2. The different part of the diagram represents the stability for the solid and soluble phases. By understanding these diagrams, it is possible to predict which redox specie that will be dominating under a given condition. (Sposito, 1998)



Figure 3-2. Pourbaix diagram for arsenic (Smedley and Kinniburgh, 2002

If the electron activity is taken into account, the constant of the reaction can be calculated in the same way as for an equilibrium reaction. By adding the reduction of a proton, the electrons are eliminated without changing the log K for the reaction. Almost all redox reactions are varieties of this general equation shown below (Sposito, 1998).

$$pC_{ox} + xH^{+} + e^{-} \leftrightarrows qC_{red} + yH_{2}O_{(1)} \qquad \text{(Equation 3-6)}$$
$$K = \frac{[C_{red}]^{q}[H_{2}O]^{y}}{[C_{ox}]^{p}[H^{+}]^{x}[e^{-}]} \qquad \text{(Equation 3-7)}$$

3.3. Arsenic

Arsenic exists in nature as As^{3+} , As^{5+} , As^{3-} and As^{0} . Under reducing conditions As(III) (arsenite) is dominating while As(V) (arsenate) is dominating under oxic conditions. Even though the arsenic atom is positively charged, it is forming anions with oxygen, and is binding to positively charged binding sites (WHO, 2001). Due to arsenates strong binding capacity to solids, it is less soluble and less mobile than arsenite (Goh and Lim, 2005).

Arsenite is not only more mobile and soluble than arsenate, it is also proved to be more toxic than arsenate (smaller amount is needed to reach a toxic response), and because of this, it is assumed that arsenite is the carcinogen. Since the arsenate is a phosphate analogue it can inhibit important processes in the body e.g. phosphorylation of ADP to ATP. The arsenite has high affinity to sulphydryl groups and binds efficiently to keratin rich tissue as hair or nails. By inhibiting important enzymes it can also create a depletion of carbohydrates. (WHO, 2001)

Geological source

The most abundant arsenic bearing mineral is arsenopyrite (Smedley and Kinniburgh, 2002). The geological source of arsenic in the Bengal area is believed to be located further upstream, in the mountain area, where the river originates (Nickson et al., 2000). The initiating process of the transport of arsenic would be the physical weathering of rocks in the Tibetan plateau and when the grain sizes is small enough, chemical weathering can take place at a significant rate. Iron silicates and arsenic bearing sulphides is believed to be the origin of arsenic (Saunders et al., 2004). When these are exposed to weathering processes, iron oxy-hydroxides are formed with arsenic adsorbed to them, and transported downstream towards the ocean. The colloidal fractions have generally a short residence time in fluvial water systems, only about 2 weeks in a major river. The larger fractions, and sand with coatings of iron oxyhydroxides, that are deposited on the river bed, has a residence time up to 1000- 10 000 years. Both sizes are good in adsorbing arsenic, but since the colloidal fractions are flushed out in to the ocean they will have no impact on arsenic concentrations in the delta. (Saunders et al., 2004;McArthur et al., 2004)

Redox based mobilisation processes

Since arsenic normally is bound to the sediments in the form of pyrite and oxyhydroxides minerals (mainly iron), some mechanism has to release it. To release arsenic from pyrite, an oxidation of pyrite has to occur, while a reduction is necessary to release the arsenic from oxyhydroxides.

Normally the groundwater is anoxic. Changing these conditions to oxic (e.g. introducing oxygen to the aquifer system) would increase the possibilities for arsenic to be released in the groundwater, (DTU, 2005) by means of oxidation of arsenopyrite, and a positive relationship between As and pe is expected.

 $4 \text{ FeAsS}_{(s)} + 13 \text{ O}_2 + 6 \text{ H}_2\text{O} \leftrightarrows 4 \text{ SO}_4^{2-} + 4 \text{ H}_3\text{AsO}_4^{3-} + 4 \text{ Fe}^{3+} + 12 \text{ H}^+$

However, McArthur et al., 2004) argue that, if this is the case, arsenic would be sorbed by FeOOH and not released in the groundwater (McArthur in DTU, 2005).

<u>The</u> most widely accepted theory today about arsenic release is the reduction of iron oxyhydroxides. When high concentrations of organic matter are present, the organic carbon is degraded by micro-organisms, and the redox conditions are changed by an increased electron activity. The electron donated by this oxidation will reduce iron oxyhydroxides present in the sediments, and solid Fe(III) is reduced to dissolved Fe^{2+} . The arsenic adsorbed to the iron oxyhydroxides in the form of arsenate will be reduced to arsenite and dissolve. Only when the oxidation of organic carbon is significant, i.e. a complete reduction of the iron oxyhydroxides, the reduction can result in the mobilisation of arsenic (Smedley and Kinniburgh, 2002; DTU, 2005).

8 FeOOH ⁺ CH₃COO + 15 H₂CO₃ \leftrightarrows 8 Fe²⁺ + 17 HCO⁻ + 12 H₂O

Even though a relationship between Fe and As could be expected, it is not necessary the case. Variations can occur due to uneven distribution of As in the minerals and resorption of As to minerals. The lack of relationship could also be due to precipitation of Fe and HCO₃ to FeCO₃, since HCO₃ is produced at the reduction of iron oxyhydroxides. (Nickson et al., 2000; McArthur et al., 2004; DTU, 2005)

In groundwater with elevated levels of arsenic, elevated concentrations of DOC are normally found (> 1 mg/l). The positive relationship that often is found indicates that peat, and peat derived DOC is contributing to the elevated concentrations of arsenic. It is however a secondary relationship, where DOC is reducing the iron oxyhydroxides that releases the arsenic bound to it (Anawar et al., 2003).

Non-redox-based mobilisation processes

Due to the similarities in size and chemical properties, arsenic typically desorb from the minerals, if an extent of phosphate is present. If this ion exchange occurs, there will be a positive relationship between phosphate and arsenic (present as arsenate).

3.4. Ammonium

Ammonium is a positively charged ion, NH_4^+ , of ammonia NH_3 . It is present in nature under reducing conditions. Normal levels in ground and surface water are typically below 0.2 mg/l, but under reducing conditions levels around 3 mg/l are not rare. Higher concentrations typically indicate some kind of contamination process, e.g. agricultural activities, industrial processes, disinfection with chloramines or sewage, or from natural degradation of peat. (WHO, 1986)

When peat layers are degraded by micro-organisms, organic N is first mineralised to ammonium. Depending on which of the redox states that is present, different types of micro-organisms are degrading the peat (i.e. which type of micro-organism can use the oxygen in the present form). According to Harms-Ringdahl (2007), the peat present in the Nam Du is unlikely to be the source of ammonium, partly because of the high C/N ratio. A high C/N ratio indicates that the peat at Nam Du is old and has been degraded for some time already and when the peat is degraded, only low concentrations of NH₄⁺ are expected. Further, the δ^{15} N in the ammonium in the groundwater is very high, especially compared to the δ^{15} N of the organic N in the peat. Typically, there is no fractionation of nitrogen isotopes when organic nitrogen is mineralised to ammonium and no enrichment of ¹⁵N is expected from this process. Therefore, the peat at Nam Du is not a likely source, and instead surface activities are believed to cause the elevated concentrations of ammonium. (Harms-Ringdahl, 2007)

4. Methods and materials

4.1. Sampling plan and laboratories

The sampling sites were chosen in order to get transect lines along the groundwater flow (see Figure 4-1). The sample codes are summarised in Table 4-1. Individual wells were chosen as starting points, sites at the riverbank and ponds were chosen to fit the direction of the groundwater flow. A sketch of a transect line and sampled mediums are shown in Figure 4-2. We identified transects and sampled at different depths in order to explore the relationship between the groundwater flow and the concentrations of As and NH_4 ions.



Figure 4-1. Map of the Nam Du area with sampling points and transects perpendicular to the river. Transects are located along the main flow direction of the groundwater.

P1AB, P3AB and P86AB	Monitoring wells
В	Upper aquifer (qh)
Α	Lower aquifer (qp)
ND3, ND 9, PY and PX	Production wells
NDT 12	Observation well used for old pumping tests
Pond 1, Pond 2 and Pond 3	Ponds surface water
1RGw, 2RGw and 3RGw	River groundwater
2RSw	River surface water
Rb1, 2, 3	River bed (for infiltration measurements)

Table 4-1. Summary of sample codes



Figure 4-2. Schematic picture of locations and sampled media along a transect at the Nam Du field site.

Due to the different types of analyses and our intentions to support collaboration with several laboratories, multiple samples were taken at each location and sent to different laboratories. The laboratories are listed in Table 4-2, and the analyses in each laboratory are summarised in Table 4-3.

Table 4-2. The laboratories used in the study.

CETASD - Centre for Environmental Technology and Sustainable Development, Hanoi
EAWAG -Swiss Federal Institute of Aquatic Science and Technology, Switzerland
INST – Institute of Nuclear Science and Technology, Hanoi
ÅAU – Åbo Akademi University, Finland
SU – Stockholm University, Sweden
SLU – Swedish University of Agricultural Sciences, Umeå

Laboratories	Main comp + DOC.	As spec.	Trace el.	$\frac{\delta^{15}N}{NH_4^+}$	$\delta^{15}N, \\ \delta^{18}O \\ NO_3^{-1}$	$\delta^{18}O, \\ \delta^{2}H \\ H_{2}O$	³ H	δ ³⁴ S	δ ¹⁵ N in org-N Sediments
EAWAG	120 + 25 ml	50 ml	50 ml						
CETASD	500 ml		500 ml						
INST	500 ml		500 ml	31	31	50 ml	1.5 1		
ÅAU								101	
SU	120 ml	50 ml	50 ml						
SLU (Umeå)									200 g

Table 4-3. A summary of all laboratories, sample type and sampled volumes.

To characterize the spatial distribution of parameters supposed to affect the fate of nitrogen and arsenic, water and sediment samples were taken at different locations and at different depths. A summary of the locations for the different analyses are given in Table 4-4.

Table 4-4. A summary over conducted analyses and the samples collected at the respective sampling sites. All
samples are water samples except Sediments. An "X" means that all sites were analysed for the given parameters,
however, some parameters were not investigated for all sites and for these parameters the sampled sites are named.

Sampling sites	Main comp.	As spec	Trace el	¹⁵ N/ ¹⁴ N NH4 ⁺	¹⁵ N/ ¹⁴ N ¹⁸ O/ ¹⁶ O NO ₃ ⁻	¹⁸ O/ ² H H ₂ O	³ H	³⁶ S	C/N Ratio	¹⁵ N/ ¹⁴ N org N
Ponds (D)	Х	X	Х	Х	Х	X				
River surface water (RSw)	X	х	х	Х	Х	Х				
Shallow groundwater (RGw)	X	х	х	RGw 1	Х	Х				
Production wells	X	X	Х	Х	Х	Х	ND9	Х		
Monitoring wells	Х	Х	Х	Х	Х	Х	PX, P86A P3A	Х		
Sediments	X ²	X ²	X^2						Х	X

In this study, no analyses of isotope studies and C/N-ratio were performed, these results are discussed in Harms-Ringdahl, 2007).





² Results not yet available

4.2. Collection of samples in the field

Surface water from the ponds and the river

The surface water samples from the ponds and river, which should be analysed at EAWAG were collected directly from the water surface at all locations. Water that should be analysed at INST or CETASD was first transferred to a bucket from which subsamples intended for the two laboratories could be collected

Shallow groundwater from the riverbank

To extract shallow groundwater at the riverbank, a partly perforated, PVC tube (diameter ~ 30 mm) with one pointed end, was pushed down about 0.5 m into the sediment, at a distance of one to three meters from the shoreline. The open upper end of the tube was sealed with sticky tape and a hose connected to a vacuum pump was

pushed through the tape. Water was extracted by creating a vacuum with the pump, (Figure 4-3). In order to minimize the amount of sediment that entered the tube, a ladies' stocking was attached over the perforated part of the tube.

Production wells

At Nam Du, the production wells were between 60-70 m deep which means that they extracted water from the lower aquifer system (qp).With one exception (PX), all of the production wells had taps from which untreated water could be collected. A hose was fitted to the tap and water samples were collected directly from the hose. At the private production well, PX, there was no tap and the pipe system was dismounted before the treatment plant. A sample could be collected from a hose fitted directly to the open pipe by sticky tape. All pumps in the production wells can be submerged. Thereby the risk of oxygen introduction was low. At the private well PY the water passed an aerated sand filter before sampling. For further information concerning PY, see see section 7.7, sources of error.

Monitoring wells

The monitoring wells, P1, P3, P86 and NDT12, were situated in transect 2 and 3, see Figure 4-1. The depth of the wells and the water level were measured with a measuring tape fitted with a solder. The "P" wells had two wells at each site which covered both aquifers. The depth of the wells varied from 20 to 70 meters depending on the aquifer; see Appendix X for exact depths. The letter A after the number indicates the lower aquifer and a B indicates the upper aquifer. Before the collection of groundwater samples three well volumes of water were extracted and removed with a generator driven pump before the samples were collected directly from the pump hose. At P86B and P1B the recharge was so slow that it was not possible to empty three volumes within the available time. These wells were totally emptied once and samples were collected on the following day. The pump was connected with a Grundfos BMI/MP1-220V to adjust the frequency/flow rate and electricity was provided by a fuel generator. The pump could be submerged and was designed to stop air entrance into the water during pumping. This was done in order not to interfere with measurements of Dissolved Oxygen and redox potential.

<u>Deep sediments</u>

Sediment samples from the aquifers were obtained by drilling. The drilling took place in transect two, close to ND 9 (Figure 4-1). A diesel driven rotating drill was used and the planned depth was 60 m. The diameter of the drill was 110 mm and that of the casing was 127 mm. The cores that could be sampled with this equipment had a diameter of 91 mm. From 0 - 5.5 m and from 33.0 - 46.0 m, soil was sampled continuously. In the sandy layer (5.5 - 33.0 m), only one sample from the uppermost part could be collected and the length of the other samples that were collected varied between 0.13 - 0.50 m. The first 5.5 meter increment was drilled without a casing but when the drill entered the sandy layer, a casing had to be installed to avoid any collapsing of sand into the bore hole. After 15 m, the casing collapsed. It was not possible to withdraw it. Instead the drilling was continued through the casing. To avoid the sand fraction to collapse into the bore hole, bentonite clay was used to stabilize the profile. At 46 m, the drill reached the coarser layer of the Hanoi aquifer (LAS) and could not continue. Because of the bentonite, the outermost layer of the core was removed before taking the actual sediment sample. From each one meter increment a sample of 200 g was collected and dried at HUMG and then sent to the SLU laboratory in

Umeå for analysis. No peat layers were found during the sampling campaign and therefore no information about the isotopes in fresh peat could be achieved. Instead we were provided with old peat from earlier drillings at Nam Du (ND16) carried out by Hanoi Drilling Company in 2003. These samples were sent to SLU Umeå for analysis of δ^{15} N. Two samples from the drilling and two from ND16 were analysed at KTH by Gunnar Jacks, for exchangeable ammonium.

Surface sediment

Samples of about 200 g bottom sediment were collected from the three sampling points at the riverbank and at the three ponds. Samples were collected at the bottom approximately one meter from the shoreline. To take the sediment bottom samples, a 30 mm pvc tube was pressed down into the sediment and about 30 cm long cores were collected and put into plastic bags that were sealed. Before drying the samples were stored in a fridge at 8 °C. The samples from the ponds and river shoreline were dried in an oven at 30 °C, until air-dry, at SGI's laboratory in Linköping. Samples from the drilling were dried at HUMG at 70 °C until dry. Twenty-five grams of dry material were sent for analysis both to Stockholm University for total analysis and to SLU, Umeå for isotope analysis of nitrogen. One sample from Pond 2 and one from Riverbed 3 were analysed at KTH by Gunnar Jacks, for exchangeable ammonium.

4.3. Analyses in the field

Analyses of pH, Eh and temperature

The pH was measured in the field with a Jenway 370 pH/mV-meter fitted with a pH epoxy electrode, see Figure 4-4.



Figure 4-4. Nguyen Thi Thanh Thuy and Johanna Moreskog is measuring pH at pond 2.

At the production and monitoring wells, a flow-through cell was used (Figure 4-5). At the ponds and the river, the measurements were done directly in the water. Regarding the

shallow groundwater at the riverbank, samples were measured in a 120 ml plastic bottle. The redox potential (Eh) was measured in the field with the same Jenway 370 pH/mV-meter fitted with an Eh electrode. To decrease the sample contact with oxygen the measurements were done in a flow-through cell. Temperature was measured with the same equipment.



Figure 4-5. The flow-through cell which was used for measuring Eh, pH and temperature.

Infiltration capacity

To estimate the infiltration rate at the riverbank (Rb1, 2 and 3), a seepage meter was used. The seepage meter was constructed of a metal can connected to a sealable plastic bag (Figure 4-6). The can was pressed into the sediments below the water surface in the riverbank and the bag was filled with 500 ml of water. The water was allowed to pass in two directions, which means that both net infiltration *in* to the riverbank and net groundwater flow *out* from the riverbank could be measured. After 1 hour, the amount of water in the plastic bag was measured and the direction of the flow and the flow rate could be calculated. If net infiltration was occurring there should be less than 500 ml water in the bag and if there was an outward flow the bag should contain more water. The surface area of the can was 509 cm^2 .



Figure 4-6. Seepage meter used for measuring the seepage at the riverbanks.

Conductivity and dissolved oxygen

The electrical conductivity (C) and dissolved oxygen (DO) were measured in the field with a WTW Multi 197i, fitted with one electrode for conductivity and one for dissolved oxygen. The conductivity and DO in surface water were measured directly in the water, whereas water from the wells was pumped into a bucket during measurements. Shallow groundwater from the riverbank could not be measured with this device since the extracted volume of water was too small. Instead Eh was measured in a small glass bottle, to give an indication of the prevailing redox conditions.

4.4. Treatment/handling of samples for laboratory analyses

The Swiss Federal Institute of Aquatic Science and Technology, Switzerland (EAWAG)

Water samples intended for the EAWAG laboratory were sucked into a 60 ml syringe. For each sample a disposable 0.45 μ m Millipore filter was fitted to the syringe and the sample was injected into an acid washed plastic bottle. For the main components and DOC, 120 + 25 ml bottles were used (140 ml was needed in the laboratory). The major component samples were stored in a fridge at 8 °C to reduce microbial activity. 50 ml acid washed plastic bottles were used for trace element analysis. Each sample was acidified immediately to pH < 2 using suprapur 1:1 HCl, and pH indicator paper was used to estimate the pH value. Arsenic fractionation was done in the field by fitting an anion exchange column to the filter and syringe. The column adsorbed As(V) and As(III) could leave the column. The samples that had passed through the column were collected and treated in the same way as the samples for trace elements. Both trace elements and arsenic speciation samples were stored at room temperature.

Centre for Environmental Technology and Sustainable Development, Vietnam (CETASD)

Samples intended for the CETASD laboratory, were collected with a 60 ml syringe. A disposable 0.45 μ m Millipore filter was fitted to the syringe and the sample was injected into 500 ml plastic bottles. The procedure was repeated until the bottles were full. One of the bottles was used to analyse main components and the other was used for trace elements. Both samples were treated and stored in the same way as the samples for EAWAG. However, water from the ponds contained such large amounts of sediments that the samples had to be filtered in the laboratory and not in the field.

Institute of Nuclear Sciences and Technology, Vietnam (INST)

The samples for nitrogen isotopic analysis of ammonium ions at the INST laboratory were collected in 3 L plastic bottles and acidified to pH <2 with HCL 1:1. For nitrogen isotope analysis of nitrate, 3 L of water were collected. A 500 ml sample was collected for analysis of main components. Another 500 ml sample was collected for analysis of trace elements and acidified to pH < 2 with HCL 1:1. For δ^{18} O and δ^{2} H (deuterium) fractionation, a 50 ml sample was collected below the water surface to prevent oxygen entering the sample. Water for the tritium (³H) analysis was collected in three 500 ml bottles. Due to the large volumes needed, only a few of the samples were filtered in the field. Most of the samples were filtered in the laboratory about one week after sampling. All samples were stored at room temperature, about 1 month, before they were analysed. The procedures for isotope analysis are further described in section 4.5.

Åbo Akademi University Finland (ÅAU)

For the sulphur isotope analysis (δ^{34} S/ 32 S), a 10 L water sample was collected in two 5 L bottles. The sample was passed through an ion exchange column to enrich the sulphate (see Figure 4-7).The columns were stored cool and brought to Sweden where the sulphate was eluted. The eluated samples were sent to Åbo for analysis. Unfortunately, the analyses did



Figure 4-7. Water passed the ion exchanger by siphon.

not succeed, either due to too small amounts of S, or incorrect treatment before sending them to ÅAU.

Stockholm University (SU)

In order to get a picture of the local conditions and to be able to construct a relevant sampling schedule, samples were sent to Sweden for immediate analysis at the Stockholm University. These samples were taken only at a few easily available locations in the beginning of the field campaign. However, due to unpredictable circumstances the results were delayed and could not be used as intended.

For sampling of water for analysis at the SU laboratory, water was sucked into a 60 ml syringe. A disposable 0.45 μ m filter was fitted to the syringe and the sample was injected to acid washed plastic bottles. For the main components 120 ml bottles were used, and for DOC 20 ml bottles. For trace elements, 50 ml acid

washed plastic bottles were used. Arsenic speciation was done in the field by fitting an anion exchange column to the filter and syringe. The sampled volume was 50 ml. Both trace elements and arsenic speciation samples were acidified immediately to pH < 2 using suprapur 1:1 HCl and pH indication paper to estimate the pH. The main components and DOC samples were stored in a fridge at 8 °C to reduce microbial activity. All the other samples were stored at room temperature.

Swedish University of Agricultural Sciences, Umeå (SLU)

Dried sediment samples from surface sampling and cores from the drilling campaign were sent to Umeå for δ^{15} N/¹⁴N analysis and for determining the carbon/nitrogen ratio.

4.5. Analyses in laboratory

In this section the procedures in the laboratories are described. Since we only had the possibility to participate in the laboratory process at CETASD and INST, only the procedures at these laboratories are included. However, at all laboratories the samples were handled and analysed by professional laboratory staff.

Nitrogen and oxygen isotopes in NO₃ at INST

The water samples were first filtered through a 0.45 μ m filter and then passed through a column packed with 2 g of Dowex AG1-X8 anion-exchange resin. Nitrate is retained by the resin and the other anions like Cl⁻ and SO₄²⁻ pass through the column. Nitrate from the column were eluated with NaCl (1M). The eluate was dried using a dry freezer in order to get NaNO₃ crystals. 610 μ g of NaNO₃ as a minimum was wrapped in a tin capsule and analyzed on a mass-spectrometer for δ^{15} N. A part of the nitrate eluate was treated with BaCl₂ to remove any sulphate associated with nitrate during the eluation. The solution was passed through a column packed with 1 g AG 50W X8 to remove the excess Ca²⁺. The obtained solution was dried with a dry freezer to get sulphate-free nitrate for δ^{18} O in NO₃ analysis in a mass-spectrometer. For results and analyse of these data see Harms-Ringdahl (2007).

*Nitrogen isotopes in NH*₄⁺ *at INST*

Samples were filtered through a 0.45 μ m Millipore filter and then passed through a column packed with 2 g AAG 50W X8 cation exchange resin to retain NH₄⁺. The enriched ammonium was eluated from the column with 10% HCl. The excess HCl was neutralised with NaOH (0.05M) to pH 6.8. The ammonium-chloride was crystallised using a dry freezer and approximately 385 μ g NH₄Cl was placed in a tin capsule for analysis of δ^{15} N in a mass-spectrometer. For results and analyse of these data see Harms-Ringdahl (2007).

Main components at INST

The sample was filtered through a 0.2 μ m filter and diluted 100 times with deionised water. A volume of 25 μ l of the sample was then analysed by ion chromatograph (IC) to determine the concentrations of an- and cations, using two different colons.

Main components at CETASD

The sample was filtered through a 0.2 μ m filter and diluted 100 times with deionised water. A volume of 25 μ l of the sample was then injected into an IC to determine the concentrations of an- and cations.

Trace elements at INST

The sample was filtered through a 0.45 μ m filter and diluted 100 times with deionised water. The filtered sample was injected into an ICP-MS and the results were displayed on a computer.

Trace elements at CETASD

20 ml of sample was injected into a plastic vial and inserted the auto sampler of an atomic adsorption spectrometer (AAS).

5. Results

5.1. Drilling log

The log from the drilling that was made is shown in Figure 5-1. Note that the LCN2 aquiclude (clay layer) is missing at the location of the drilling. It should have appeared between D and E in the figure. This indicates that there are hydraulic windows between the aquifers and that the sediment layers are not continuous and of different thickness.



Figure 5-1. The drilling log from the borehole next to ND 9. The hydrogeological units are named according to the general hydrogeological formations. Note that all layers, except for LCN 2, are present.

5.2. Results from the field analyses

The results from the field measurements, pH, conductivity (C), dissolved oxygen (DO), temperature (T) and redox (pe), are presented in Table 5-1. The measured pH is ranging from 6.29 - 8.69, but is overall around neutral. All surface water samples have a pe above 5, except for sampling point 3RGw. The high pe is correlating well with log DO (see Figure 5-2). Two groups can be distinguished; (1) surface water with higher pe, implying oxidising conditions, and (2) groundwater with lower pe and reducing conditions.

The infiltration decreased downstream in the Red River from 2.4 l/m^2h to 0.4 l/m^2h between transect 1 and 2. In transect 3 it was an outflow of 1.2 l/m^2h to the Red River from the aquifer.

Sampling site	Param	eters to meas	ure in field			
~F	pН	C (µS/cm)	DO (mg/l)	DO (%)	T (°C)	ре
Pond 1	8.21	509	9.5	115	25.1	5.41
Pond 2	7.40	325	4.48	52.7	23.7	5.78
Pond 3	7.54	309	5.45	65.4	24.8	5.95
ND3	6.65	495	0.08	0.9	26.5	1.23
ND9	6.35	555	0.15	2	27	1.98
1RGw	7.08	NA	NA	NA	23.9	6.61
2RGw	7.30	NA	NA	NA	24.6	NA
2RSw	8.00	188	6.86	80.6	23.8	6.27
3RGw	8.69	NA	NA	NA	16.7	3.65
P1A	7.11	1068	0.25	3	27.4	0.88
P1B	6.68	692	0.12	1.4	26.5	1.34
P3A	6.29	572	0.15	1.9	27.4	1.57
P3B	6.29	777	NA	NA	24.5	1.66
PX	6.31	381	0.33	3.5	27.2	2.86
P86A	6.95	774	0.2	2.5	26.2	1.45
P86B	6.90	635	NA	NA	24.4	2.13
ND12-4(A,C)	7.10	294	0.21	2.7	25.9	1.47

Table 5-1. Summary of the results from the field measurements of pH, conductivity (C), dissolved oxygen (DO), temperature (T) and redox (pe).



Figure 5-2. There is a relationship between Log DO and pe. LAS – Lower Aquifer System. UAS – Upper Aquifer System.

5.3. Chemical analyses

The results presented here are based on analyses made at EAWAG unless stated otherwise. The samples analysed at EAWAG are assumed to be the most representative due to the small sampling volume, i.e.: (1) the same standard procedure could be applied at all locations, and (2) the samples could be filtered in the field without delay. This is further discussed in section 7.7.

Table 5-2 summarises some important parameters from the laboratory analyses together with pH and pe from the field measurements. It can be observed that sulphate and nitrate is only present in surface waters and shallow groundwater close to the river. Ammonium is, except for two sampling locations, always more abundant than nitrate. Arsenic, PO_4 , Fe and Mn is mostly present in groundwater where the pe is low.

Sampling	As(tot)	As(III)	As(V)	Fe	Mn	SO ₄ -	NH ₄	NO ₃	PO ₄	DOC	HCO ₃ -	ре	pН
site	μg/l	μg/l	μg/l	mg/l	mg/l	mg/l	mg/l	mg/l	mg/l	mg/l	mg/l		
Pond 1	4.9	NA	NA	0.1	0.0	40.0	2.1	0.0	0.0	9.4	211	5.41	8.21
Pond 2	2.6	NA	NA	0.0	0.5	24.0	0.5	0.0	0.0	5.2	160	5.78	7.40
Pond 3	1.9	NA	NA	0.1	0.0	19.0	3.1	1.6	0.0	6.1	147	5.95	7.54
ND3	83.0	63.0	20.0	9.2	0.5	0.0	5.5	0.0	2.7	2.6	332	1.23	6.65
ND9	83.0	77.0	6.0	11.7	0.5	0.0	11.1	0.0	1.4	2.7	349	1.98	6.35
1 RGw	2.6	0.9	1.7	0.1	0.1	6.6	0.6	0.0	0.0	1.7	139	6.61	7.08
2 RGw	0.4	0.2	0.2	0.0	0.0	16.0	0.0	0.0	0.0	1.9	220	NA	7.30
2 RSw	2.8	NA	2.8	0.2	0.0	9.5	0.0	1.4	0.0	2.0	109	6.27	8.00
3 RGw	2.6	1.8	0.8	0.0	0.0	24.0	0.5	2.7	0.0	3.3	101	3.65	8.69
P1A	257.0	258.0	0	10.6	0.4	0.0	16.4	0.0	2.9	4.0	450	0.88	7.11
P1B	264.0	254.0	10.0	12.6	0.8	0.0	44.7	0.0	0.6	10.5	696	1.34	6.68
P3A	152.0	148.0	4.0	19.3	0.1	0.0	30.9	0.0	3.2	12.2	253	1.57	6.29
P3B	39.0	34.0	5.0	37.3	0.5	0.0	12.6	0.0	2.2	9.0	338	1.66	6.29
РХ	27.0	27.0	0.0	18.7	0.7	0.0	4.1	0.0	0.7	2.1	222	2.86	6.31
P86A	312.0	282.0	30.0	7.0	0.4	0.0	11.8	0.0	2.2	2.9	467	1.45	6.95
P86B	62.0	56.0	6.0	6.0	0.6	0.0	3.3	0.0	0.0	2.4	412	2.13	6.90
ND12-4	44.0	39.0	5.0	3.5	1.1	0.0	2.3	0.0	0.0	2.0	190	1.47	7.10

Table 5-2. Selected results from laboratory analyses and field measurements..

As can be seen in Figure 5-3 below, dissolved arsenic increases as pe decreases. Further, Figure 5-4 shows that arsenic is primarily abundant as arsenite, As(III).



Figure 5-3. A negative relationship between pe and arsenic can be observed. LAS – Lower Aquifer System.

Figure 5-4. Most of the total arsenic is present as arsenite, according to the laboratory analyses.

UAS – Upper Aquifer System.

There does not seem to be any relationship between arsenic and iron, see Figure 5-5, nor does DOC seem to relate to Arsenic, see Figure 5-6. However, it should be noted that P3A is differing from the other samples from LAS regarding the DOC concentrations, see Figure 5-6. Phosphate (PO₄) was plotted against Arsenic but no relationship could be found (figure not shown). Arsenic has a positive relationship to HCO₃, particularly within the aquifers (Figure 5-7), and a weak positive relationship to ammonium (Figure 5-8). The sample location P3A, is deviating from the other samples in the LAS regarding NH₄⁺.





Figure 5-5. There seems to be no relationship between As and Fe. LAS – Lower Aquifer System. UAS – Upper Aquifer System.

Figure 5-6. No relationship can be seen between DOC and As. P3A is differing from the other in LAS.



Figure 5-7. A relationship between As and HCO₃ can be seen.



Figure 5-8. There seems to be no relationship between arsenic and ammonium for the different aquifers in particular. P3 A is differing from the other in LAS.

Ammonium seems to have a positive relationship both to HCO_3 and DOC (Figure 5-9 and Figure 5-10), but a negative relationship with pe (Figure 5-12). However, no relationship could be seen between DOC and pe (Figure 5-11), nor between DOC and HCO_3 (Figure 5-13). Finally, there seems to be a negative relationship between HCO_3 and pe (Figure 5-14). In Figures 5-9 to 5-11 and 5-14, the result from P3A seems to deviate due to the high concentration of ammonium and DOC found in this location.



Figure 5-9. A relationship can be seen between NH₄ and HCO₃. P3A is deviating from the other samples in the LAS. LAS - Lower Aquifer System. UAS -Upper Aquifer System.



Figure 5-11. No relationship can be observed between pe and DOC. P3A is deviating from the other samples in the LAS.



Figure 5-13 No relationship can be observed between Figure 5-14 A negative relationship between pe and DOC and HCO₃. P3A is deviating from the other samples in the LAS.



Figure 5-10. A relationship can be seen between NH₄ and DOC. P3A is deviating from the other samples in the LAS.



Figure 5-12. A negative relationship between pe and NH₄ can be seen.



HCO₃ can be seen.

5.4. Comparisons between the laboratories

Figures 5-15 to 5-18 present some typical diagrams from the comparison of chemical data between the laboratories. In general, the results from EAWAG and CETASD are similar, and the results from INST are deviating somewhat. Data mainly from EAWAG was used, partly due to the comparison but also since a larger number of parameters were analysed at EAWAG than at CETASD and INST. For further information, see the discussion in section 7.7.



Figure 5-15. The difference in measured total arsenic concentrations from the three laboratories. EAWAG is set as standard against which INST and CETASD are compared.



Figure 5-16. The difference in measured iron concentrations from the three laboratories. EAWAG is set as standard against which INST and CETASD are compared.



Figure 5-17. The difference in measured nitrate concentrations from the three laboratories. EAWAG is set as standard against which INST and CETASD are compared.



Figure 5-18. The difference in measured ammonium concentrations from the three laboratories. EAWAG is set as standard against which INST and CETASD are compared.

6. Geochemical modelling in Visual Minteq 2.51

Visual Minteq (VM) is a chemical equilibrium model and is developed as a Windows version of MINTEQA2. The model can be used to study ion speciation, solubility, ion-exchange, and to calculate the speciation of redox-couples and metal-humic complexations. In this study, VM 2.51 and the standard databases included with VM 2.51 were used (Gustafsson, 2007b). Two types of calculations were made using VM: (1) distribution of arsenic species as a function of different assumed redox conditions, and (2) solubility calculations in order to investigate different minerals solubility index (SI). When modelling with VM, one run with a certain input data can give several outputs. In this case, only one run is necessary for both calculations mentioned above.

Before importing the input data for the equilibrium calculations, the redox couples were set (As, Mn, Fe, S) and pH was fixed. Even if measured concentrations of DOC in the input data were used when running the model, 5 mg/l DOC (Nica-Donnan) had to be added to the model after instruction from the programmer (Gustafsson, 2007a). The input data used are pH, temperature, pe and the total concentrations of each anions, cations and trace elements (detailed input data can be found in Appendix A). For the model with pe measured in field, all samples were included, but since redox processes is a minor process in surface waters, these data were excluded from further models with changed pe.

6.1. Redox and speciation in VM

VM was used to investigate how the field measurements of pe and the chemical analyses of Arsenic species relate to each other. When the pe and pH of each sample is plotted in a stability diagram for different arsenic compounds, most of the arsenic should exist as arsenate, As(V), see Figure 6-1. However, as presented in the previous section, Arsenic is mostly present as arsenite, i.e. As(III), see Figure 5-4. Measuring redox in the field is difficult, and therefore VM was used to investigate at which pe the measured distribution of As(III) and As(V) is likely to appear. In order to do this, the model was run for a number of different pe-values, but with the rest of the input data constant.



Figure 6-1. A stability diagram of different arsenic compounds compared to measured pH and pe. According to this stability diagram, most of the arsenic is present as arsenate.

The distribution between the arsenic species, according to the modelling in VM was investigated for every sampling location. In Figure 6-2 and Figure 6-3 (ND 9 and P86 B) two rather typical results from the modelling can be seen, representing both aquifer systems. The diagrams include the measured As-speciation, in order to understand how the model relates to the measured pe. In Figure 6-2, it can be observed that As(V) and As(III) has inverted values in the field, compared to those calculated with VM for the measured pe. That is, As(III) is more abundant than arsenate in the field, but should, according to the model, be less abundant. The measured values of the arsenic species, rather seems to correspond with the model at a pe-value around 0, instead of the measured value around pe 2. In Figure 6-3 the measured values of arsenite and arsenate were approximately the same at pe 2, which according to the model, would be more likely around a pe value of 0. However, at pe 0, the measured concentrations are also 100 times lower, than estimated by the model around pe 0.



Figure 6-2. Arsenic species at ND 9 according to VM at different pe, together with the measured arsenic species.



Figure 6-3. Arsenic species at P86 B according to VM at different pe, together with the measured arsenic species.

According to the model, the ferrous (Fe^{2+}) ion is always dominating over the ferric (Fe^{3+}) in all groundwater samples at all pe values. Since ferrous iron will dissolve, a high concentration of iron can be expected in the water. In all surface waters samples, the ferric ion dominates over the ferrous. No iron in solution should thus be expected in the surface water, since ferric iron will precipitate under normal conditions. Further, Mn^{2+} is always dominating over Mn^{3+} in all water samples and at all pe values used in the model. Ammonium is dominating over nitrate, including surface waters, at most points (except 2 RSw, 3 RGw and 2 RGw).

At all pe and in all sampling points, SO_4^{2-} is dominating over HS⁻, except for when pe is lowered with 5 units, see example in Figure 6-4. At pe -5, HS⁻ is dominating in all samples except for P86B, PX and ND12.

Figure 6-4. Sulphur species at P1 A according VM at different pe.

According to the model, sulphate is not reduced during the present field conditions, and should not be excepted until pe is lowered by at least 3.5 units. However, iron and manganese are reduced but most likely not the sulphate. The reduction sequence of the principal elements including Arsenic is shown in Figure 6-4. From the modelling it can be concluded that the pe in the aquifer should be where arsenic is typically reduced, i.e. between that of iron and sulphate reduction. The interval at which the groundwater in Nam Du is likely to be characterised by, is marked out with red in Figure 6-5.

Figure 6-5. The reduction sequence of the principal elements including arsenic. The interval at which the groundwater in Nam Du is likely to be characterised by, is marked out with red

6.2. Solubility calculations in VM

It is also of interest to investigate if minerals are over- or under saturated, i.e. if they are likely to precipitate or not. The precipitation of minerals controls to which extent a substance is likely to be found in dissolved phase, e.g. arsenic. If the arsenic bearing minerals are likely to be dissolved, it is likely to find arsenic in solution. However, since no studies of which minerals are actually present in Nam Du were available, these results are entirely based on the modelling in Visual Minteq. Minerals that, according to the model, are oversaturated in all water samples are; Al(OH)₃ (soil), Boehmite, Diaspore, Gibbsite (C) and Kaolin. This means that they should precipitate and thus could be present in solid phase. Minerals that are oversaturated in all groundwater samples are presented in Table 6-1. There is a domination of Fe and Al minerals, but also some silicates are present. The only arsenic bearing mineral present, according to the model is BaHAsO₄:2H₂O, which according to the model, is not present in all locations. The mineral is more stable in samples with higher concentrations of dissolved arsenic. When modelling with a pe of 1 unit lower than that measured in the field, the stability of the mineral decreased a lot, and the mineral would only be stable where there are very high concentrations of arsenic (P1A-B, P86 A). The stability of some iron oxyhydroxides decreases with decreasing pe, see Table 6-1 (for stoichiometry, see Appendix B).

Table 6-1. The minerals which are oversaturated according to model in Visual Minteq at all samples sites at different pe values. Since BaHAsO₄:2H₂O is the only arsenic bearing mineral present, this mineral is also included, even though it only is oversaturated at some spots.

			ł	ре		
Minerals	field	-1	-2	-3	-4.25	-5
Al(OH) _{3 (Soil)}	х	х	х	х	х	х
Boehmite	х	х	x	х	х	х
CoFe ₂ O ₄	х	х	x	х	х	х
Diaspore	х	х	x	х	х	х
Fe(OH)2.7Cl ₃	х	х	х		х	х
Ferrihydrite	х	х				
Ferrihydrite (aged)	х	х				
Gibbsite (C)	х	х	x	х	х	х
Goethite	х	х	х	х		
Halloysite	х	х	x	х	х	х
Hematite	х	х	x	х	х	
Hercynite	х	х	x	х	х	х
Imogolite	х	х	x	х	х	х
Kaolin	х	х	x	х	х	х
Lepidocrocite	х	х	х	х		
Maghemite	х					
Magnesioferrite	х					
Magnetite	х	х	х	х	х	х
Quartz	х	х	х	х	x	х
Siderite	х	х	х	х	х	х
BaHAsO ₄ :2H ₂ O	х	х				

7. Discussion

7.1. *Redox conditions*

The measured values of pe and the fact that DO could be measured in the samples indicate that the groundwater is slightly oxidised. However, this is contradicted by the dominance of arsenite in the analysed samples and the geochemical modelling in VM. The deviation is probably explained by introduction of oxygen during sampling, since only very little oxygen is needed to contaminate the sample and increase the pe. The positive relationship between DO and pe (which was found) is also expected, since pe should increase when oxygen is introduced to the system. Lack of relationship could indicate an error in one, or both, of the methods.

Instead, the results indicate that the groundwater in Nam Du is reduced. It is shown that iron is reduced and in solution. The absence of sulphate could be explained either by a reduction of sulphate or due to that there simply is no sulphate. According to the results from VM, SO_4^- is not expected to be reduced unless pe is at least 3.5 units lower than that measured. Further, if the absence of sulphate was due to a reduction to HS⁻, it would not be possible to find any arsenate, or at least very little, since arsenate is reduced before sulphate. Since as much as 30 µg/l of arsenate is found at some sites it is not likely that the sulphate is totally reduced. More likely is that dissolved sulphate is absent in this samples, either it doesn't exist at all or it is bound to solid phase.

7.2. Solubility and precipitation of minerals

Minerals that, according to the model, are oversaturated in all water samples are $Al(OH)_{3(soil)}$, Boehmite, Diaspore, Gibbsite(C) and Kaolin. All these minerals are also constituents of bauxite, a prolific rock present in Vietnamese geological formations. Bauxite has also been studied for its properties of adsorbing arsenite and arsenate with the purpose of cleaning water, which proved to be very successful (Bhakat et al., 2006; Bhakat et al., 2007). This raises the question how good these minerals are to bind arsenic in nature and if they can act as a sink for arsenic, since they are stable under more reducing conditions than the iron oxyhydroxide.

The stability of some iron oxyhydroxides (e.g. ferrihydrite and goethite, see Table 6-1) seems to decrease when pe decreases, which can have effect on the arsenic mobilisation. At pe from field, both ferrihydrite and goethite is oversaturated. Since these are two common iron oxyhydroxides (Pedersen et al., 2006), they might have an arsenic load. If pe measured in field would be representative for the environment in the aquifers, most of the iron oxyhydroxides would be precipitated, in solid phase. When pe is decreased, both ferrihydrite and goethite is dissolved according to the model in VM, and sorbed arsenic can dissolve. As already discussed in section 6.1, it is likely that pe measured in field is slightly overestimated, possibly due to introduction of oxygen during the sampling.

Due to the lack of measured data on the minerals that can be expected in the Red River delta, it is difficult to conclude which of the minerals above that actually are present in the Nam Du sediments. However, big reservoirs of kaolin are present northeast of Hanoi (see section 2.4) and thus, at least kaolin is likely to be present. This is not the case in Bangladesh, where instead very little kaolin is present (Akai et al., 2004).

Since the concentrations of phosphate are rather low and the redox conditions are low (arsenite is dominating), no competitive ion exchange is expected. According to the models with pe values from field measurements, siderite and magnetite is oversaturated in all groundwater samples except. Vivianite, is oversaturated at all points except for the sample points ND12 and P86B. However, chemical analyses from these sites showed no measurable concentrations of phosphate in the groundwater and since vivianite is a phosphate mineral this might be why it is absent in the model.

7.3. *Mobilisation process of arsenic*

The two main mobilisation processes of arsenic are: (1) mobilisation of arsenic due to the reduction and subsequent dissolution of iron-hydroxides, and (2) mobilisation of arsenic due to the oxidation of Arsenopyrite.

The low levels of sulphate, the relatively high pe, pH around 7 and the results from the model where sulphate is not reduced, indicate that no oxidation of sulphurous mineral occurs. Furthermore, if arsenic would be released by oxidation of arsenic bearing sulphurous minerals, the relationship between arsenic and pe would be positive instead of the observed negative relation. In addition, low concentrations of sulphate in the water and in the bedrock and a low abundance of sulphurous minerals is reported in previous studies (Berg et al., 2001). Thus, oxidation of arsenic bearing sulphurous minerals is *not* likely to be the main process mobilising arsenic.

Instead, under the prevailing reducing conditions, several processes are creating HCO₃ as a product and a relationship between HCO₃ and both arsenic and ammonium were observed in this study. Peat is abundant at several locations in the sediments in Nam Du. When the organic matter is degraded in the aquifer, the environment becomes more reducing and the arsenic bearing iron oxyhydroxides are reduced and consequently mobilising the arsenic (Nickson et al., 2000; Smedley and Kinniburgh, 2002). The lack of relationship between arsenic and iron can be explained by precipitation of iron and HCO₃, e.g. siderite, vivianite and magnetite (Sracek et al., 2004). Since HCO₃ has several sources during reducing conditions, the total concentration of HCO₃ will not be as affected of this precipitation as the concentrations of iron are expected to be (McArthur et al., 2004; Sracek et al., 2004). This study indicates, and confirms previous studies (e.g. Berg et al., 2001), that reductive dissolution of iron oxyhydroxides might be one of the major processes releasing arsenic to the groundwater. This is also the most accepted theory regarding arsenic release in the groundwater in Bangladesh.

7.4. Ammonium and the origin of DOC

Geochemical modelling in VM indicates that ammonium is dominating over nitrate, at most points, including surface waters and it would not persist in its reduced form unless there were reducing conditions. Since the surface waters are oxidised, the source of ammonium has to be continuous, so that the nitrogen can persist in the reduced form, ammonium. As long as the source is continuous, it is possible for ammonium to persist, since the transformation of nitrogen is an internal process in microbes, and not dependent of redox changes in the same way as e.g. Fe. Using e.g. C/N-ratio and isotope studies, Harms-Ringdahl (2007) concluded that the ammonium in the groundwater in Nam Du does not derive from the peat layers in the sediments, but that surface activities is a more likely source. Further, there is no clear relationship between DOC and pe, which might suggest that the DOC in the aquifer does not derive from the degradation of peat. A relationship can, on the other hand, be seen between DOC and NH₄, which might indicate that the DOC reducing the iron oxyhydroxides is from the same source as the ammonium. As ammonium is not believed to be derived from the peat, ammonium would probably not relate to DOC if the DOC was a product from the peat. When the Fe-redox couple governs the redox level, there is no drive for oxidation of ammonium, which might be why arsenic and ammonium is correlating well.

7.5. Mobilisation of arsenic and ammonium

With no detectable levels of sulphate, the relatively high pe, pH around 7 and the results from the model (where sulphate is not reduced) indicate that no oxidation of sulphurous mineral occurs. Further, if arsenic would be released by oxidation of arsenic bearing sulphurous minerals, the relationship between arsenic and pe would be positive instead of the observed negative relationship. In addition, low concentrations of sulphate in the water and in the bedrock and a low abundance of sulphurous minerals is reported in previous studies (Berg et al., 2001). Thus, oxidation of arsenic bearing sulphurous minerals is not likely to be the main process mobilising arsenic.

With the prevailing reducing conditions several processes are creating HCO₃ as a product and a relationship between HCO₃ and both arsenic and ammonium were observed in this study. When the organic matter is degraded in the aquifer, the environment becomes more reducing and the arsenic bearing iron oxyhydroxides are reduced and mobilising the arsenic (Nickson et al., 2000; Smedley and Kinniburgh, 2002). The lack of relationship between arsenic and iron can be explained by precipitation of iron and HCO₃ e.g. siderite, vivianite and magnetite (Sracek et al., 2004). Since HCO₃ has several sources during reducing conditions, the total concentrations will not be as affected of this precipitation as the concentrations of iron are expected to be (McArthur et al., 2004; Sracek et al., 2004). This study indicates, and confirms previous studies (e.g. Berg et al., 2001), that reductive dissolution of iron oxyhydroxides might be one of the major processes releasing arsenic to the groundwater. This is also the most accepted theory regarding arsenic release in the groundwater in Bangladesh.

Geochemical modelling in VM indicates that ammonium is dominating over nitrate, at most points, including surface waters and it would not persist in its reduced form unless there were reducing conditions. Since the surface waters are oxidised, the source of ammonium has to be continuous, so that the nitrogen can persist in the reduced form, ammonium. This is possible since the redox processes of nitrogen is known to be a rather slow process.

Using e.g. C/N-ratio and isotope studies, Harms-Ringdahl (2007) concluded that the ammonium in the groundwater does not derive rfom peat, but that surface activities are a more likely source. Even if the ammonium does not derive from the peat layers, it is still possible that the peat layers are causing a change in the redox conditions. The degradation of peat can mobilise DOC which can be further transported downwards in the profile and create reducing conditions and thus mobilising arsenic due to reduction of iron oxyhydroxides. However, the lack of relationship between DOC and pe, suggests that the

DOC might not derive from redox processes, and degradation of peat. However, it can also be due to a secondary relationship with pe. A relationship can be seen between DOC and NH₄, which might indicate that the DOC reducing the iron oxyhydroxides is from the same source as ammonium. And as ammonium is not derived from peat, it would probably not relate to DOC if it were a product from peat. The fact that arsenic and ammonium is correlating well, suggesting there might be some common ground for mobilisation.

Finally, the observed levels of ammonium are not likely to be a direct health problem, since ammonium itself rarely has any hazardous effects. However, it is often used as an indicator for contamination of anthropogenic waste and pollutants associated with it. The arsenic levels on the other hand are well above both international and local guidelines (10 μ g/l). Water distributed via the treatment plant, is aerated and not likely to cause any health problem, since it will precipitate with iron. The problem is likely the uncontrolled private wells, where no standard for cleaning is set.

7.6. *Health issues*

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7.7. Sources of error

Measuring the redox potential in groundwater is problematic, since it is difficult to reach the aquifer without disturbing the anaerobic environment. The electrodes easily get polluted by oxygen which is reproduced into the result. Introducing oxygen in the groundwater sample will increase the redox potential and be misguiding. In reality a reaction also rarely reaches equilibrium, which means that it is very difficult to obtain reliable values of the redox potential comparable to existing equilibrium constants, which calculations and models often are based on. The safest way to measure the redox potential in groundwater, even though no procedure is prominent, is to install the electrode in the aquifer, or at the bottom of the well. But this is difficult and demanding. The next best thing to do would be to measure the redox potential in a flow trough cell, with this the introduction of oxygen is decreased, and also a constant flow is created around the electrode. (Back, 2001) Since it was not possible to use the flow through cell at all locations due to low flow (i.e. shallow groundwater and surface waters), the different methods might cause unrealistic relationship between the sampling points. The surface water was expected to be oxidised and was of less interest and it was studied more as an indication of redox condition, rather than the actual redox potential.

One of the wells, PY, was a private well for drinking water supply, and an aeration filter had been installed which could not be passed by during the measurements. This means that the samples collected were aerated and did not correspond to the actual conditions in the aquifer at the location. When the sampling was performed, oxygen was most probably introduced, and most dissolved iron would have been oxidized to Fe^{3+} and precipitated.

Since the sample was filtered, precipitated iron which might have arsenic adsorbed to it, would have been removed from the sample.

The fact that no peat was observed during the drilling might be because of the rotary drill that was used. In Ravenscroft et al. (2001) it is stated that such drill might veil the presence of peat.

Since CETASD and INST needed larger sample volumes than EAWAG, some of the samples had to be filtered in laboratory since the high content of particles made the filtering process slow. This was the case for all samples from the ponds to CETASD and all samples to INST, except for samples from ND 3 and ND 9. With the filters, small particles and bacteria could be removed. When delaying this procedure the chemical composition of the sample is likely to change when the conditions are changed from anoxic to oxic.

The differences between the laboratories (shown in section 5.4) can however, not only be explained by delayed filtering. Different laboratories have different procedures for analysing samples, e.g. EAWAG did their analyses of arsenic with MS, while CETASD used a HG-AAS and INST used IC. That CETASD and EAWAG are correlating well with each other can be explained by the fact that CETASD is a Vietnamese laboratory supported by EAWAG, and the two laboratories thus have similar procedures. The samples to EAWAG were also smaller which makes them easier to handle and to avoid oxygen contamination. However, these samples were sent by mail and the samples were probably not kept cold during transport. Note that total nitrogen levels also differ between the laboratories which can not be explained by the filtering but rather with the different analysis methods. Another possibility could be denitrification, i.e. that nitrogen escapes as N_2 gas, but as the samples were acidified and kept in air tight bottles this should not happen.

8. Conclusions

Below, the main conclusions drawn from this study are listed.

- Geochemical modelling in Visual Minteq together with the measured As-species indicates a slightly more reducing environment than what the redox measurements in the field indicate.
- The negative relationship between arsenic and pe, the lack of sulphate and the results from the geochemical modelling indicate that the oxidation of arsenic-containing sulphurous minerals is not likely to be the main process to mobilise arsenic.
- The results from this study support the current hypothesis that the mobilisation of arsenic is due to the reduction of iron hydroxides, since there is a negative relationship between As and pe.
- The lack of relationship between Fe and As, can be explained by precipitation to FeCO₃.

9. Future studies and recommendations

Some questions have emerged during this study which might be of interest for future studies:

- The origin of the DOC in the aquifer should be investigated since the DOC is an important factor for the mobilisation of arsenic. This study could not clarify whether the DOC is deriving from the degradation of the peat, or if it has the same source as ammonium, thus possibly anthropogenic. An investigation of coliform bacteria could indicate if there is a surface source.
- A sequential leaching test together with detailed information about the mineral composition in the Nam Du well field would provide useful information about the arsenic association with the minerals in the sediments as an input to geochemical modelling.
- Previous studies (Berg et al., 2001) have not found any relationship between dissolved arsenic and sorbed arsenic. Since this study is entirely based on analyses of water samples, analyses of sediment bound arsenic would be of great value in order to understand the relationship between dissolved and sorbed arsenic.
- A different drilling method should be considered for coming investigations of the peat occurrence in the sediments.
- Studies of sulphate isotopes would be of great value to investigate whether the lack of sulphate is due to reduction to HS⁻ or not.

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Appendix

	Fe	Mn	Na	K	Ca	Mg	As(tot)
Sampling site	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	µg/L
	ICP-OES	ICP-OES	ICP-OES	ICP-OES	ICP-OES	ICP-OES	MS
1 DSw	0.08	0.02	28.8	20.2	45	9.8	4.9
2 DSw	0.03	0.46	14.4	7.3	41	9.5	2.6
3 DSw	0.06	0.03	17.8	6.80	40.00	6.90	1.90
ND3	9.2	0.5	6.9	3.20	68.00	21.50	83.00
ND9	11.7	0.47	17.5	4.40	50.00	27.30	83.00
1 RGw	0.08	0.07	4.1	2.10	37.00	5.40	2.60
2 RGw	0	0	4.3	2.90	63.00	9.40	0.40
2 RSw	0.2	0	4.1	1.20	30.00	5.70	2.80
3 RGw	0	0	4.4	1.50	31.00	5.90	2.60
P1A	10.6	0.38	23.5	4.60	75.00	28.10	257.00
P1B	12.6	0.84	32	7.40	99.00	42.70	264.00
P3A	19.3	0.05	37.7	4.10	19.00	13.30	152.00
P3B	37.3	0.45	27.3	1.80	73.00	24.10	39.00
PX	18.7	0.74	17.3	4.30	27.00	17.60	27.00
PY	0	1.02	58.2	1.80	6.40	6.80	0.40
P86A	7	0.36	23.6	3.90	104.00	30.10	312.00
P86B	6	0.57	11.2	1.40	92.00	22.40	62.00
ND12	3.5	1.09	4.5	2.00	40.00	10.00	44.00
	As(III)	As(V)	Sr	Ва	Li	Al	Со
Sampling site	As(III) µg/L	As(V) µg/L	Sr μg/L	Ba μg/L	Li µg/L	Al μg/L	Co μg/L
Sampling site	As(III) μg/L MS	As(V) µg/L calculated	Sr µg/L ICP-OES	Ba μg/L ICP-OES	Li µg/L MS	Al μg/L MS	Co μg/L MS
Sampling site	As(III) µg/L MS	As(V) µg/L calculated	Sr µg/L ICP-OES 135	Ba μg/L ICP-OES 124	Li µg/L MS 0.6	Al µg/L MS 30	Co μg/L MS 0.48
Sampling site 1 DSw 2 DSw	As(III) μg/L MS	As(V) μg/L calculated	Sr μg/L ICP-OES 135 138	Ba μg/L ICP-OES 124 81	Li µg/L MS 0.6 6.4	Al μg/L MS 30 25	Co μg/L MS 0.48 0.18
Sampling site 1 DSw 2 DSw 3 DSw	As(III) μg/L MS	As(V) μg/L calculated	Sr μg/L ICP-OES 135 138 114.00	Ba μg/L ICP-OES 124 81 99.00	Li μg/L MS 0.6 6.4 0.70	Al μg/L MS 30 25 38.00	Co μg/L MS 0.48 0.18 0.00
Sampling site 1 DSw 2 DSw 3 DSw ND3	As(III) μg/L MS 63.00	As(V) µg/L calculated 20.00	Sr μg/L ICP-OES 135 138 114.00 317.00	Ba μg/L ICP-OES 124 81 99.00 231.00	Li µg/L MS 0.6 6.4 0.70 2.30	Al μg/L MS 30 25 38.00 237.00	Co μg/L MS 0.48 0.18 0.00 0.45
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9	As(III) μg/L MS 63.00 77.00	As(V) µg/L calculated 20.00 6.00	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00	Li μg/L MS 0.6 6.4 0.70 2.30 4.20	AI μg/L MS 30 25 38.00 237.00 16.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw	As(III) μg/L MS 63.00 77.00 0.90	As(V) μg/L calculated 20.00 6.00 1.70	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00	Li μg/L MS 0.6 6.4 0.70 2.30 4.20 0.30	Al μg/L MS 30 25 38.00 237.00 16.00 99.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw	As(III) μg/L MS 63.00 77.00 0.90 0.20	As(V) μg/L calculated 20.00 6.00 1.70 0.20	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.00
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw	As(III) μg/L MS 63.00 77.00 0.90 0.20	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.00 0.00 0.00
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 3 RGw	As(III) μg/L MS 63.00 77.00 0.90 0.20 1.80	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80 0.80	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00 96.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00 31.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60 0.30	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00 217.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.00 0.00 0.00
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 3 RGw P1A	As(III) μg/L MS 63.00 77.00 0.90 0.20 1.80 258.00	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80 0.80 0.00	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00 98.00 96.00 408.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00 31.00 228.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60 0.30 3.70	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00 217.00 12.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.00 0.00 0.00 0.00 0.00
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B	As(III) μg/L MS 63.00 77.00 0.90 0.20 1.80 258.00 254.00	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80 0.80 0.80 0.00 10.00	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00 96.00 408.00 555.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00 31.00 228.00 399.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60 0.30 3.70 1.50	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00 217.00 12.00 16.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.00 0.00 0.00 0.00 8.90
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A	As(III) μg/L MS 63.00 77.00 0.90 0.20 1.80 258.00 254.00 148.00	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80 0.80 0.00 10.00 4.00	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00 98.00 96.00 408.00 555.00 203.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00 31.00 228.00 399.00 417.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60 0.30 3.70 1.50 6.20	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00 217.00 12.00 16.00 54.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.00 0.00 0.00 0.00 0.00 8.90 1.70
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B	As(III) µg/L MS 63.00 77.00 0.90 0.20 1.80 258.00 254.00 148.00 34.00	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80 0.80 0.00 10.00 4.00 5.00	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00 96.00 408.00 555.00 203.00 213.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00 31.00 228.00 399.00 417.00 287.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60 0.30 3.70 1.50 6.20 2.70	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00 217.00 12.00 16.00 54.00 10.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.00 0.00 0.00 0.00 0.00 0.0
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B PX	As(III) µg/L MS 63.00 77.00 0.90 0.20 1.80 258.00 254.00 148.00 34.00 27.00	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80 0.80 0.00 10.00 4.00 5.00 0.00	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00 98.00 96.00 408.00 555.00 203.00 213.00 191.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00 31.00 228.00 399.00 417.00 287.00 205.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60 0.30 3.70 1.50 6.20 2.70 5.50	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00 217.00 12.00 16.00 54.00 10.00 13.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.00 0.00 0.00 0.00 0.00 8.90 1.70 0.78 0.00
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B PX PY	As(III) μg/L MS 63.00 77.00 0.90 0.20 1.80 258.00 254.00 148.00 34.00 27.00 0.20	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80 0.80 0.00 10.00 4.00 5.00 0.00 0.20	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00 96.00 408.00 555.00 203.00 213.00 191.00 55.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00 31.00 228.00 399.00 417.00 287.00 205.00 24.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60 0.30 3.70 1.50 6.20 2.70 5.50 12.30	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00 217.00 12.00 16.00 54.00 10.00 13.00 11.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.0
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B PX PY P86A	As(III) μg/L MS 63.00 77.00 0.90 0.20 1.80 258.00 254.00 148.00 34.00 27.00 0.20 282.00	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80 0.80 0.00 10.00 4.00 5.00 0.00 0.20 30.00	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00 96.00 408.00 555.00 203.00 213.00 191.00 55.00 519.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00 31.00 228.00 399.00 417.00 287.00 205.00 24.00 218.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60 0.30 3.70 1.50 6.20 2.70 5.50 12.30 2.20	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00 217.00 12.00 16.00 54.00 10.00 13.00 11.00 8.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.0
Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B PX PY P86A P86B	As(III) µg/L MS 63.00 77.00 0.90 0.20 1.80 258.00 254.00 148.00 34.00 27.00 0.20 282.00 56.00	As(V) μg/L calculated 20.00 6.00 1.70 0.20 2.80 0.80 0.00 10.00 4.00 5.00 0.00 0.20 30.00 6.00	Sr μg/L ICP-OES 135 138 114.00 317.00 344.00 108.00 184.00 98.00 96.00 408.00 555.00 203.00 213.00 191.00 55.00 519.00 292.00	Ba μg/L ICP-OES 124 81 99.00 231.00 292.00 58.00 104.00 106.00 31.00 228.00 399.00 417.00 287.00 205.00 24.00 218.00 175.00	Li µg/L MS 0.6 6.4 0.70 2.30 4.20 0.30 0.50 0.60 0.30 3.70 1.50 6.20 2.70 5.50 12.30 2.20 0.70	AI μg/L MS 30 25 38.00 237.00 16.00 99.00 24.00 239.00 217.00 12.00 12.00 16.00 54.00 10.00 13.00 11.00 8.00 6.00	Co μg/L MS 0.48 0.18 0.00 0.45 0.11 0.00 0.0

A. Input data for the model in Visual Minteq 2.51

	Ni	Zn	Pb	Hg	U	Cr	Cd
Sampling site	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L
	MS	MS	MS	MS	MS	MS	MS
1 DSw	1.9	20	1.2	0.37	2.2	0.16	0.02
2 DSw	1.2	4.9	2.8	0.6	0.6	0.22	0.01
3 DSw	2.50	25.00	3.80	1.20	1.23	3.3	0.11
ND3	1.50	23.00	0.50	0.51	0.01	0.62	0.02
ND9	1.50	23.00	0.66	1.00	0.00	0.12	0.01
1 RGw	0.71	7.80	3.00	1.00	0.58	0.38	0.15
2 RGw	0.46	7.20	1.20	0.66	0.86	0.1	0.02
2 RSw	0.84	16.00	0.42	0.88	0.46	0.7	0.01
3 RGw	0.41	11.00	3.40	0.19	0.50	0.27	0.02
P1A	0.76	6.30	1.30	0.27	0.00	0.13	0.02
P1B	52.00	7.90	1.60	0.61	0.83	0.16	0.09
P3A	3.90	15.00	2.50	0.56	0.04	0.41	0.02
P3B	3.00	14.00	2.50	0.20	0.03	0.5	0.03
PX	0.39	165.00	3.00	0.17	0.00	0.1	0.02
PY	2.90	8.20	3.20	0.41	0.03	0.09	0.1
P86A	0.74	5.00	1.50	0.35	0.00	0.59	0.02
P86B	50.00	13.00	1.70	0.33	0.46	0.11	0.04
	0.40	4 40	4 40	0.00	0.00	0.40	0.01
ND12	0.40	4.40	1.40	0.29	0.22	0.13	0.01
ND12	0.46 PO4-P	4.40 PO4-P	1.40 Si	0.29 HCO3-	0.22 Cl-	SO4-	NO3
ND12 Sampling site	PO4-P mg/L	4.40 PO4-P mg/L	Si mg/L	HCO3- mg/L	0.22 Cl- mg/L	SO4- mg/L	NO3 mg/L
ND12 Sampling site	PO4-P mg/L Photo	4.40 PO4-P mg/L	Si mg/L AuA	U.29 HCO3- mg/L AuA	CI- mg/L AuA	SO4- mg/L AuA	NO3 mg/L
ND12 Sampling site	PO4-P mg/L Photo 0	4.40 PO4-P mg/L 0	1.40 Si mg/L AuA 3.9	U.29 HCO3- mg/L AuA 211	0.22 Cl- mg/L AuA 27.6	0.13 SO4- mg/L AuA 40	NO3 mg/L
ND12 Sampling site 1 DSw 2 DSw	PO4-P mg/L Photo 0 0	4.40 PO4-P mg/L 0 0	1.40 Si mg/L AuA 3.9 5.5	0.29 HCO3- mg/L AuA 211 160	0.22 Cl- mg/L AuA 27.6 14.7	0.13 SO4- mg/L AuA 40 24	0.01 NO3 mg/L 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw	0.46 PO4-P mg/L Photo 0 0 0	4.40 PO4-P mg/L 0 0 0	1.40 Si mg/L AuA 3.9 5.5 2.2	0.29 HCO3- mg/L AuA 211 160 147	0.22 Cl- mg/L AuA 27.6 14.7 11.6	SO4- mg/L AuA 40 24 19	0.01 NO3 mg/L 0 0 1.593592
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3	0.46 PO4-P mg/L Photo 0 0 0 0.88	4.40 PO4-P mg/L 0 0 0 2.698244	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5	0.29 HCO3- mg/L AuA 211 160 147 332	0.22 Cl- mg/L AuA 27.6 14.7 11.6 2.2	0.13 SO4- mg/L AuA 40 24 19 0	0.01 NO3 mg/L 0 0 1.593592 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9	0.46 PO4-P mg/L Photo 0 0 0.88 0.46	4.40 PO4-P mg/L 0 0 0 2.698244 1.410446	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8	0.29 HCO3- mg/L AuA 211 160 147 332 349	0.22 Cl- mg/L 27.6 14.7 11.6 2.2 8	0.13 SO4- mg/L AuA 40 24 19 0 0	0.01 NO3 mg/L 0 0 1.593592 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw	0.46 PO4-P mg/L Photo 0 0 0.88 0.46 0	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4	0.29 HCO3- mg/L AuA 211 160 147 332 349 139	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6	0.13 SO4- mg/L AuA 40 24 19 0 0 6.6	0.01 NO3 mg/L 0 0 1.593592 0 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw	0.46 PO4-P mg/L Photo 0 0 0.88 0.46 0 0	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0 0	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220	CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16	0.01 NO3 mg/L 0 0 1.593592 0 0 0 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw	0.46 PO4-P mg/L Photo 0 0 0 0.88 0.46 0 0 0 0	4.40 PO4-P mg/L 0 0 0 2.698244 1.410446 0 0 0	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5	0.01 NO3 mg/L 0 0 1.593592 0 0 0 0 0 0 1.37226
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw	0.46 PO4-P mg/L Photo 0 0 0.88 0.46 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	4.40 PO4-P mg/L 0 0 0 2.698244 1.410446 0 0 0 0 0	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8 4.2	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109 101	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7 2.7	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5 24	0.01 NO3 mg/L 0 0 1.593592 0 0 0 0 1.37226 2.700253
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A	0.46 PO4-P mg/L Photo 0 0 0.88 0.46 0 0.46 0 0.95	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0 0 0 0 0 2.912877	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8 4.2 17.8	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109 101 450	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7 2.7	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5 24 0	0.01 NO3 mg/L 0 0 1.593592 0 0 0 0 0 0 1.37226 2.700253 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B	PO4-P mg/L Photo 0 0 0 0.88 0.46 0 0 0 0 0 0 0 0 0 0 0.95 0.19	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0 0 0 0 2.912877 0.582575	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8 4.2 17.8 12.5	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109 101 450 696	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7 2.7 3.5	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5 24 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0.01 NO3 mg/L 0 0 1.593592 0 0 0 0 0 0 1.37226 2.700253 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A	0.46 PO4-P mg/L Photo 0 0 0.46 0.46 0 0.46 0 0.46 0 0.46 0 0.46 0 0.46 0 0.46 0 0.46 0 0.46 0 0.46 0 0.95 0.19 1.05	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0 0 0 0 2.912877 0.582575 3.219496	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8 4.2 17.8 12.5 15.8	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109 101 450 696 253	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7 10.3 5.8 30.2	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5 24 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0.01 NO3 mg/L 0 0 1.593592 0 0 0 0 0 1.37226 2.700253 0 0 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B	0.46 PO4-P mg/L Photo 0 0 0.88 0.46 0 0.46 0 0.47 0.95 0.19 1.05 0.71	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0 0 0 0 2.912877 0.582575 3.219496 2.176993	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8 4.2 17.8 12.5 15.8 27.7	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109 101 450 696 253 338	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7 10.3 5.8 30.2 41.9	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5 24 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0.01 NO3 mg/L 0 0 1.593592 0 0 0 0 0 1.37226 2.700253 0 0 0 0 0 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B PX	0.46 PO4-P mg/L Photo 0 0 0 0 0.88 0.46 0 0 0.88 0.46 0 0 0.10 0.95 0.19 1.05 0.71 0.24	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0 0 0 2.912877 0.582575 3.219496 2.176993 0.735885	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8 4.2 17.8 12.5 15.8 27.7 19.7	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109 101 450 696 253 338 222	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7 10.3 5.8 30.2 41.9 5.1	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5 24 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0.01 NO3 mg/L 0 0 1.593592 0 0 0 0 0 0 1.37226 2.700253 0 0 0 0 0 0 0 0 0 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B PX PY	0.46 PO4-P mg/L Photo 0.95 0.19 1.05 0.71 0.24 0	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0 0 0 2.912877 0.582575 3.219496 2.176993 0.735885 0	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8 4.2 17.8 12.5 15.8 27.7 19.7 17.9	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109 101 450 696 253 338 222 159	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7 10.3 5.8 30.2 41.9 5.1 14.6	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5 24 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 8	0.01 NO3 mg/L 0 0 1.593592 0 0 0 0 0 0 1.37226 2.700253 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B PX PX PY P86A	0.46 PO4-P mg/L Photo 0 0 0.88 0.46 0 0.46 0 0.46 0 0.46 0 0.46 0 0.46 0 0.46 0 0.95 0.19 1.05 0.71 0.24 0 0.71	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0 0 0 2.912877 0.582575 3.219496 2.176993 0.735885 0 2.176993	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8 4.2 17.8 12.5 15.8 27.7 19.7 17.9 14.8	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109 101 450 696 253 338 222 159 467	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7 10.3 5.8 30.2 41.9 5.1 14.6 16.2	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5 24 0 0	NO3 mg/L 0 0 1.593592 0 0 0 0 0 1.37226 2.700253 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
ND12 Sampling site 1 DSw 2 DSw 3 DSw ND3 ND9 1 RGw 2 RGw 2 RGw 2 RGw 2 RSw 3 RGw P1A P1B P3A P3B P3A P3B PX PY P86A P86B	0.46 PO4-P mg/L Photo 0 0 0.88 0.46 0 0.88 0.46 0 0.19 1.05 0.71 0.24 0	4.40 PO4-P mg/L 0 0 2.698244 1.410446 0 0 2.912877 0.582575 3.219496 2.176993 0.735885 0 2.176993 0	1.40 Si mg/L AuA 3.9 5.5 2.2 16.5 19.8 7.4 3.5 4.8 4.2 17.8 12.5 15.8 27.7 19.7 17.9 14.8 13.7	0.29 HCO3- mg/L AuA 211 160 147 332 349 139 220 109 101 450 696 253 338 222 159 467 412	0.22 CI- mg/L AuA 27.6 14.7 11.6 2.2 8 2.6 2.5 2.7 10.3 5.8 30.2 41.9 5.1 14.6 16.2 10.1	0.13 SO4- mg/L AuA 40 24 19 0 6.6 16 9.5 24 0 <td>NO3 mg/L 0 0 1.593592 0 0 0 0 0 1.37226 2.700253 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0</td>	NO3 mg/L 0 0 1.593592 0 0 0 0 0 1.37226 2.700253 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0

Sampling site	NH4 mg/L	DOC mg/L AuA	рE	Temp (ºC)	рH
1 DSw	2.06057	9.4	5.40906	25.1	8.21
2 DSw	0.528021	5.2	5.780933	23.7	7.4
3 DSw	3.090855	6.1	5.949966	24.8	7.54
ND3	5.537781	2.6	1.233942	26.5	6.65
ND9	11.07556	2.7	1.977688	27	6.35
1 RGw	0.579535	1.7	6.609195	23.9	7.08
2 RGw	0	1.9		24.6	7.3
2 RSw	0	2	6.271129	23.8	8
3 RGw	0.515142	3.3	3.651116	16.7	8.69
P1A	16.35577	4	0.878972	27.4	7.11
P1B	44.68861	10.5	1.335362	26.5	6.68
P3A	30.90855	12.2	1.572008	27.4	6.29
P3B	12.62099	9	1.656525	24.5	6.29
PX	4.121139	2.1	2.85666	27.2	6.31
PY	0.386357	2.4	5.848546	26.2	6.46
P86A	11.84828	2.9	1.453685	26.2	6.95
P86B	3.348426	2.4	2.129817	24.4	6.9
ND12	2.318141	2	1.470588	25.9	7.1
		KORR			
Sampling site	-4.25	-3	-2	-1	-5
	рE	рE	рE	рE	рE
1 DSw	5.40906	5.40906	5.40906	5.40906	5.40906
2 DSw	5.780933	5.780933	5.780933	5.780933	5.780933
3 DSw	5.949966	5.949966	5.949966	5.949966	5.949966
ND3	-3.01606	-1.76606	-0.76606	0.233942	-3.76606
ND9	-2.27231	-1.02231	-0.02231	0.977688	-3.02231
1 RGw	6.609195	6.609195	6.609195	6.609195	6.609195
2 RGw					
2 RSw	6.271129	6.271129	6.271129	6.271129	6.271129
3 RGw	3.651116	3.651116	3.651116	3.651116	3.651116
P1A	-3.37103	-2.12103	-1.12103	-0.12103	-4.12103
P1B	-2.91464	-1.66464	-0.66464	0.335362	-3.66464
P3A	-2.67799	-1.42799	-0.42799	0.572008	-3.42799
P3B	-2.59348	-1.34348	-0.34348	0.656525	-3.34348
PX	-1.39334	-0.14334	0.85666	1.85666	-2.14334
PY	1.598546	2.848546	3.848546	4.848546	0.848546
P86A	-2.79632	-1.54632	-0.54632	0.453685	-3.54632
P86B	-2.12018	-0.87018	0.129817	1.129817	-2.87018
ND12	-2.77941	-1.52941	-0.52941	0.470588	-3.52941

B. Results from Visual Minteq

Minerals	Stoichiometry				
Al(OH) _{3 (Soil)}	1. Al^{+3}	3. H ₂ O	-3. H ⁺	_	
Boehmite	-3. H ⁺	1. Al^{+3}	2. H ₂ O		
CoFe ₂ O ₄	1. Co^{+2}	2. Fe^{+3}	4. H ₂ O	-8 H ⁺	
Diaspore	-3. H ⁺	1. Al^{+3}	2. H ₂ O		
Fe(OH)2.7Cl ₃	-2.7. H ⁺	1. Fe^{+3}	2.7. H ₂ O	3. Cl ⁻	
Ferrihydrite	1. Fe^{+3}	3. H ₂ O	- 3. H ⁺		
Ferrihydrite (aged)	1. Fe^{+3}	-3 . H ⁺	3. H ₂ O		
Gibbsite (C)	1. Al^{+3}	3. H ₂ O	- 3. H ⁺		
Goethite	1. Fe^{+3}	2. H ₂ O	- 3. H ⁺		
Halloysite	2. Al^{+3}	2. H_4SiO_4	1. H ₂ O	-6. H ⁺	
Hematite	2. Fe^{+3}	3. H ₂ O	-6. H ⁺		
Hercynite	-8. H ⁺	1. Fe+2	2. Al^{+3}	4. H ₂ O	
Imogolite	2. Al^{+3}	$1. H_4 SiO_4$	3. H ₂ O	-6. H ⁺	
Kaolin	2. Al^{+3}	2. H_4SiO_4	1. H ₂ O	-6. H ⁺	
Lepidocrocite	-3. H ⁺	1. Fe^{+3}	2. H ₂ O		
Maghemite	-6. H ⁺	2. Fe^{+3}	3. H ₂ O		
Magnesioferrite	-8. H ⁺	1. Mg_{1}^{2+}	2. Fe^{+3}	4. H ₂ O	
Magnetite	-8. H ⁺	2. Fe^{+3}	1. Fe^{+2}	4. H ₂ O	
Quartz	1. H ₄ SiO ₄	-2. H ₂ O			
Siderite	1. Fe^{+2}	1. CO_3^{2-}			
BaHAsO ₄ :2H ₂ O	1. Ba ⁺²	1. AsO_4^{-3}	$1. \mathrm{H}^+$	1. H ₂ O	

Table 1. The stoichiometry of minerals presented in Table 6-1.